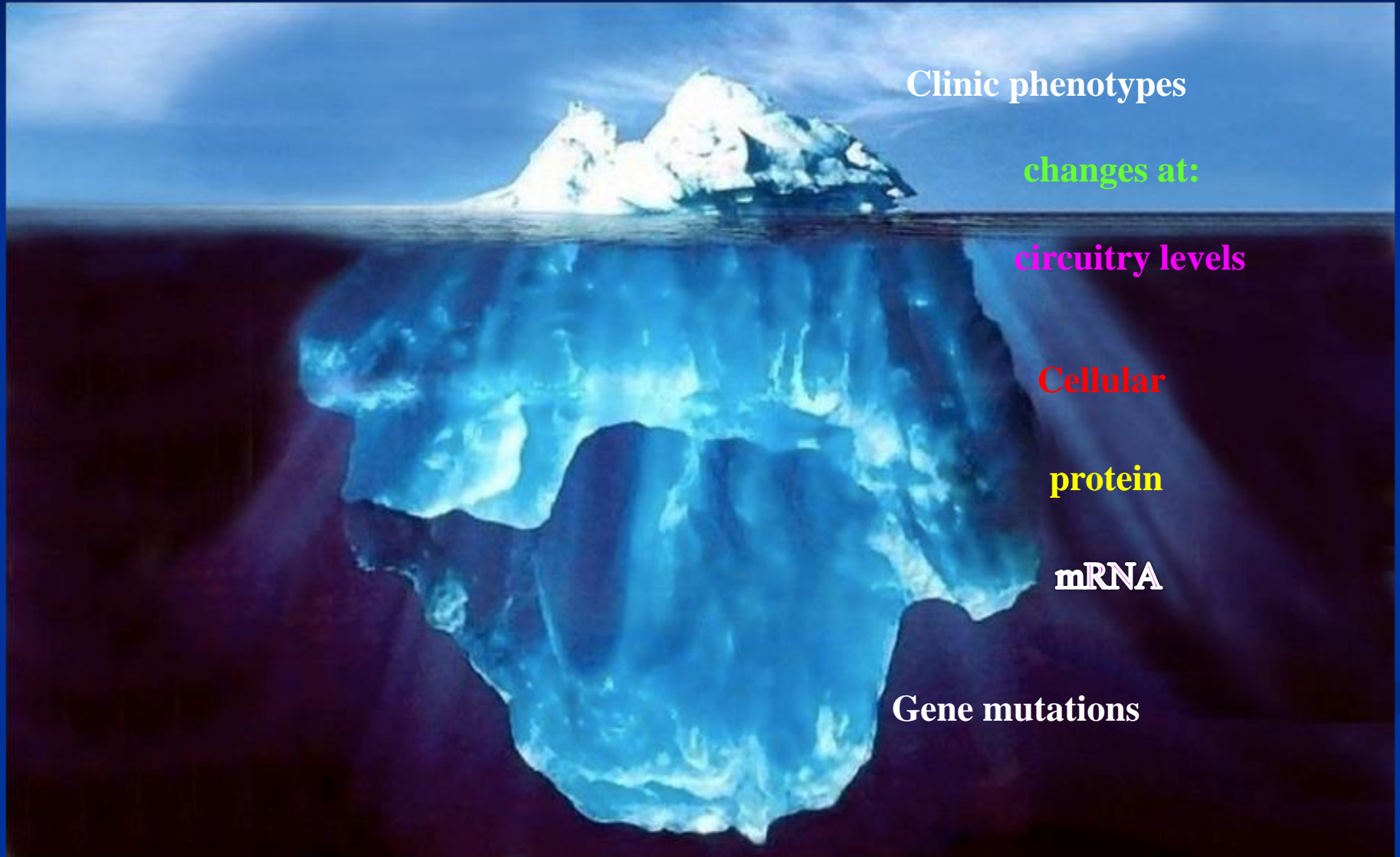


# ***SLC6A1* mutations in cell and mouse models**

**Jing-Qiong (Katty) Kang, MD, PhD  
Department of Neurology,  
Vanderbilt Brain Institute  
Vanderbilt University Kennedy Center of Human  
Development  
Vanderbilt University Medical Center  
Nashville, TN, 37232**



# What is epilepsy?



Clinic phenotypes

changes at:

circuitry levels

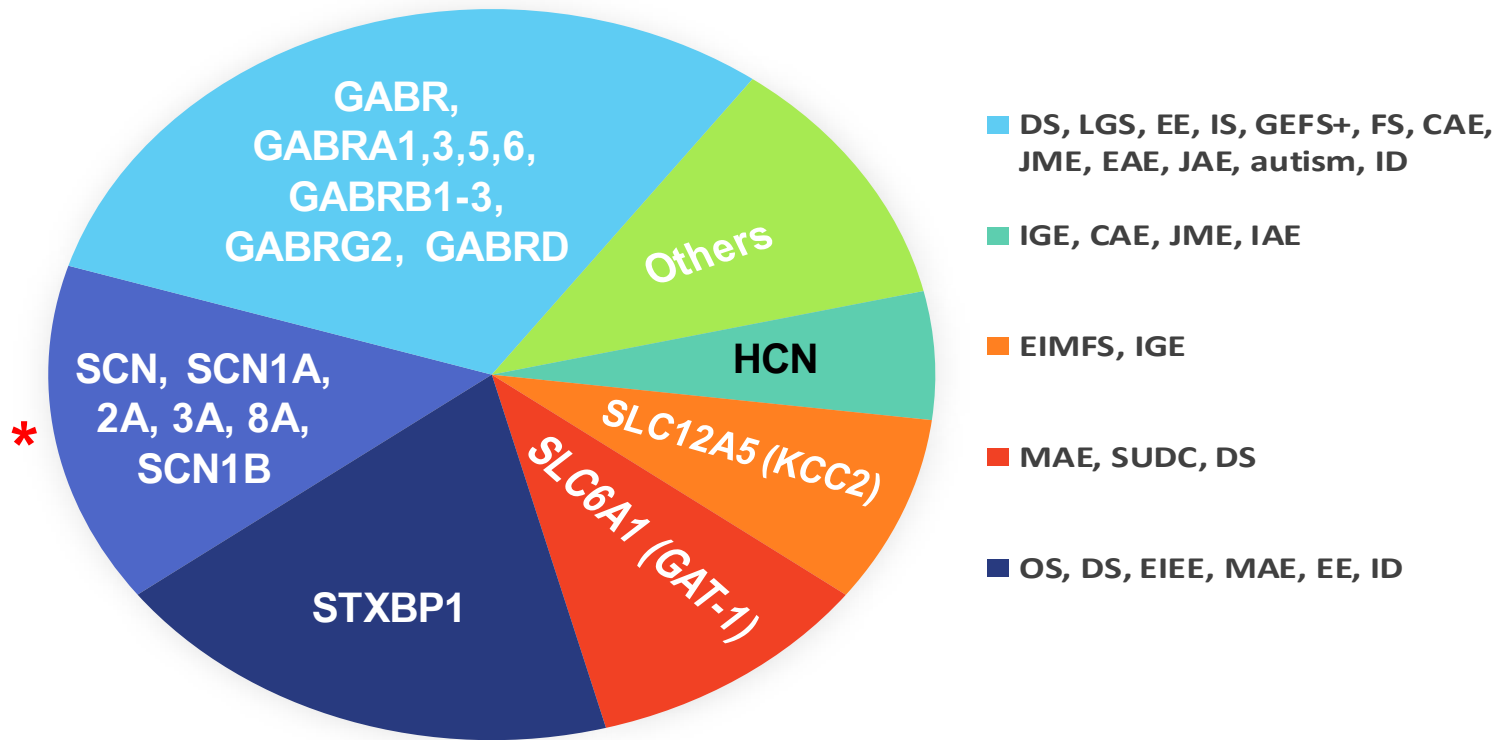
Cellular

protein

mRNA

Gene mutations

## Mutations in genes proposed in the study and the associated epilepsy syndromes



CAE=Childhood absence

DS=Dravet syndrome

EE=Epileptic encephalopathy

EIEE=Early infantile epileptic encephalopathy

EIMFS=Epilepsy of infancy with migrating focal seizures

EOME=Early onset myoclonic encephalopathy

FS=Febrile seizures

GEFS+=Generalized epilepsy with febrile seizures

IAE=Idiopathic absence epilepsy

ID=Intellectual disability

IGE=Idiopathic generalized epilepsy

IS= Infantile spasms

JAE=Juvenile absence epilepsy

JME=Juvenile myoclonic epilepsy

Lennox-Gastaut syndrome

LGS=Lennox-Gastaut syndrome

MAE=Myoclonic astatic epilepsy

OS=Ohtohara syndrome

SUDC=Sudden unexplained death in childhood

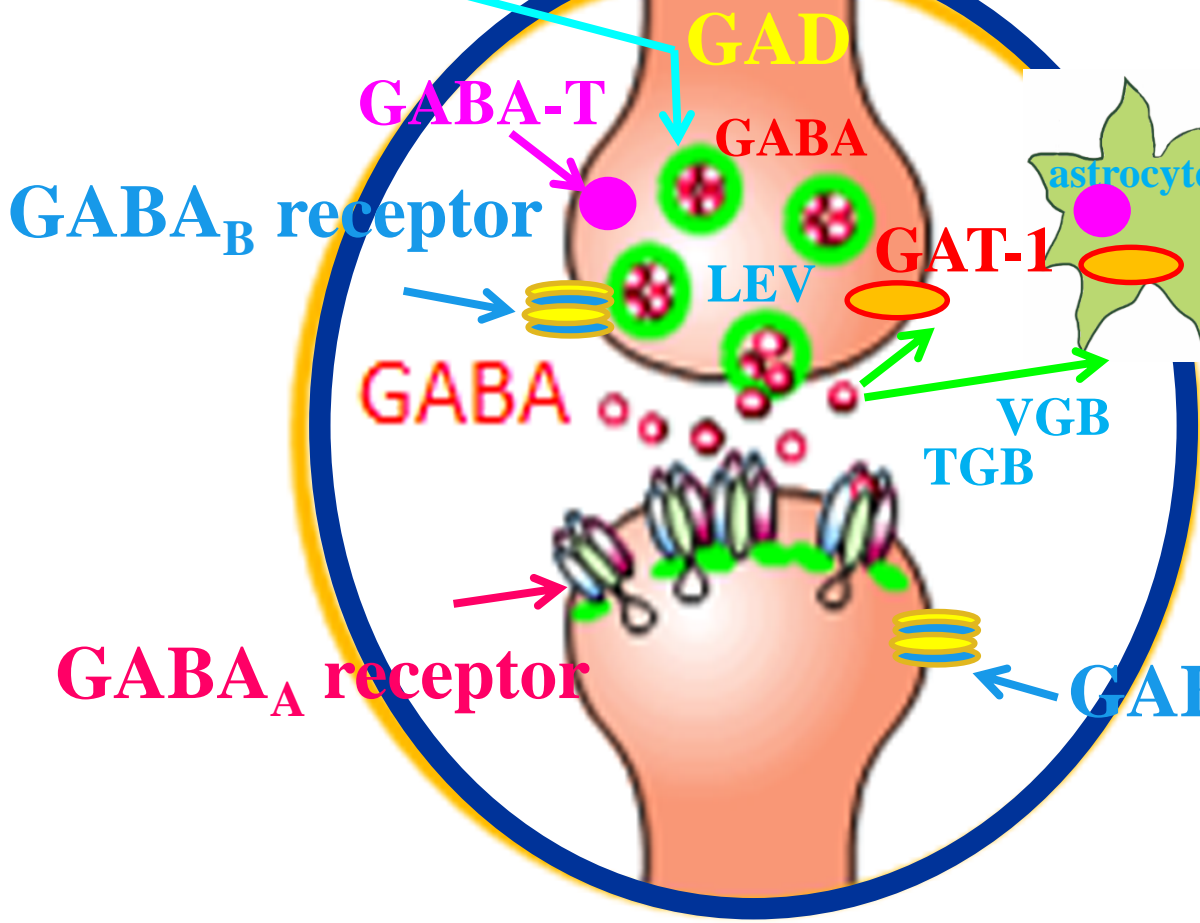
\* We will not study SCN

# GABAergic neurotransmission is a major pathway for epilepsy

## GABAergic interneuron

Receptor,  
neurotransmitter  
transporter

Glutamic acid



Antiseizure drugs

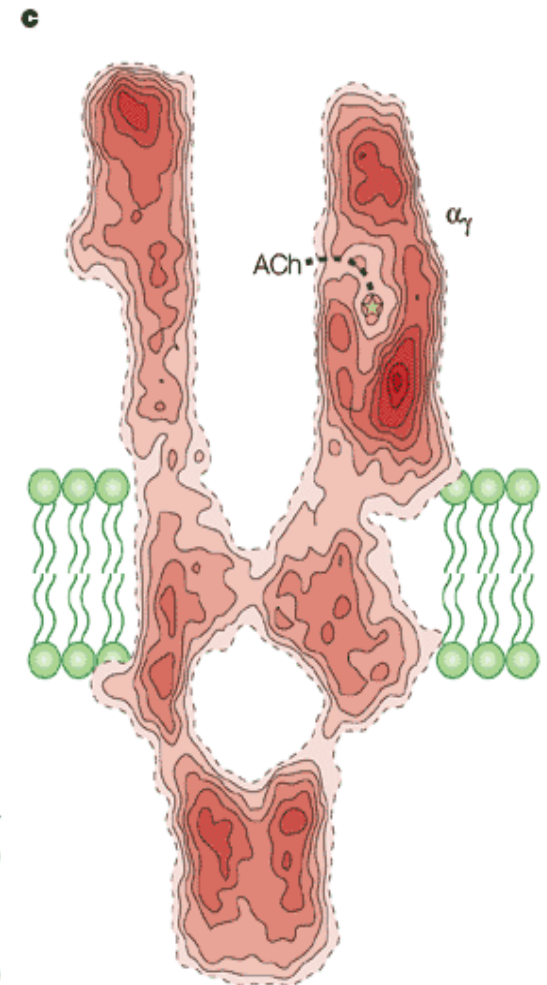
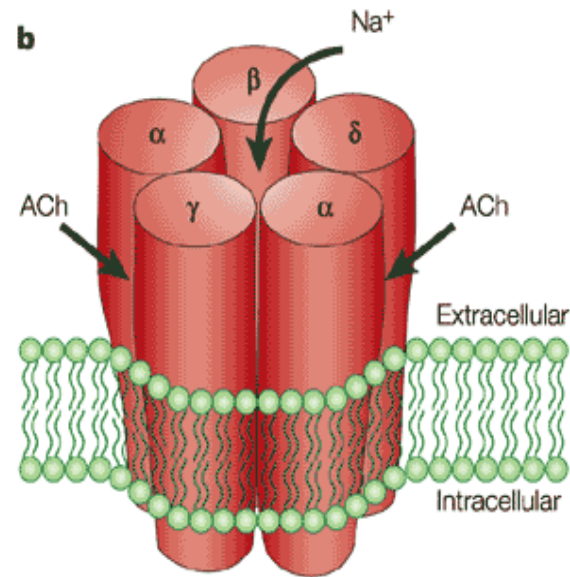
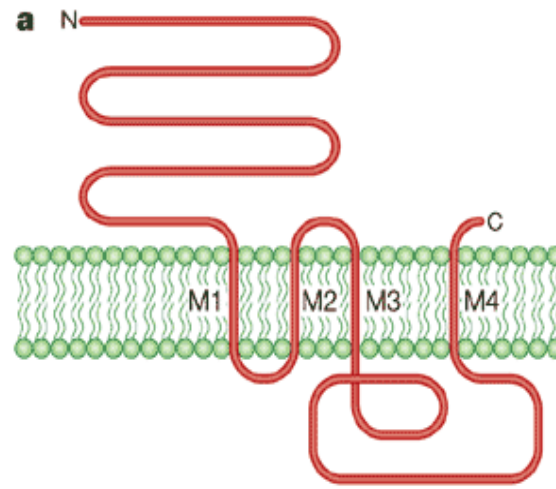
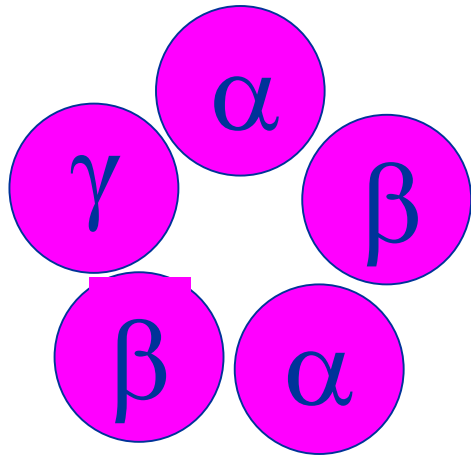
- Vigabatrin (VGB)
- Tiagabine (TGB)
- Levetiracetam (LEV)
- Clobazam (CBZ)
- Clonazepam (CZP)
- Phenobarbital (PB)

GABA<sub>A</sub> receptor

GABA<sub>B</sub> receptor

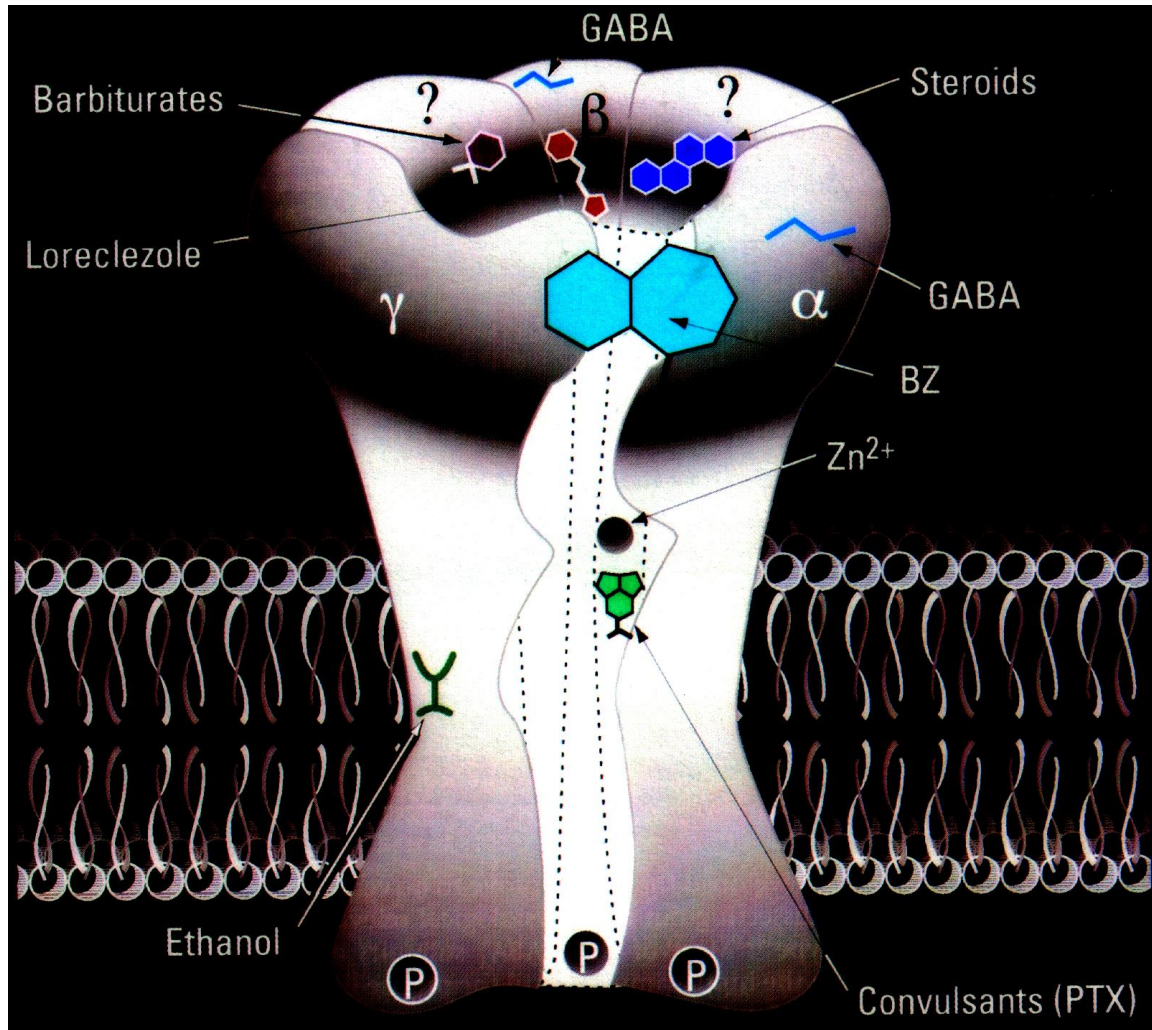
Post-synaptic neuron

# Our previous work has mainly focused on GABA<sub>A</sub> receptor



Many sedative, anticonvulsant drugs work by modulating GABA<sub>A</sub> receptors

GABA<sub>A</sub> receptor binding sites



# Our research focused on catastrophic epilepsies, ~30% resistant to current AEDS

Genes have been identified to be related to  
epilepsy in GABA<sub>A</sub> receptors and  
GABA transporters

## Receptors

GABRG2

GABRA1

GABRB2

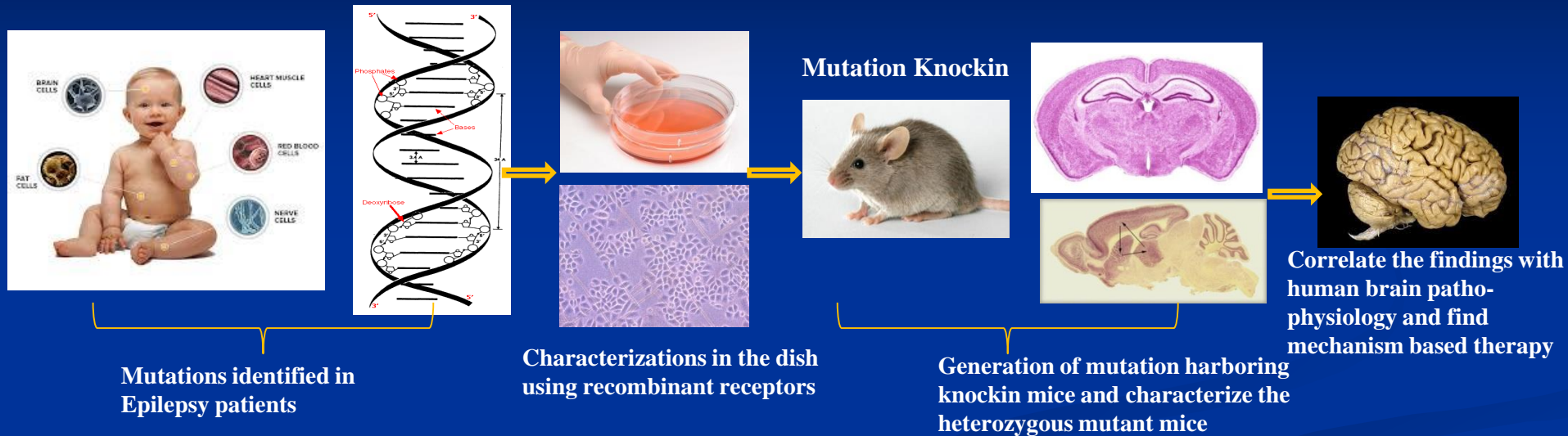
GABRB3

## Transporter

GAT-1

# Our research

## We try to identify mechanism-based therapies for genetic epilepsies



## Ongoing research

GABRA1 knockout

GABRG2 knockout

GABRB3 knockout

GABRA1(A322D) knockin : JME

GABRG2(Q390X) knockin: GEFS+/DS

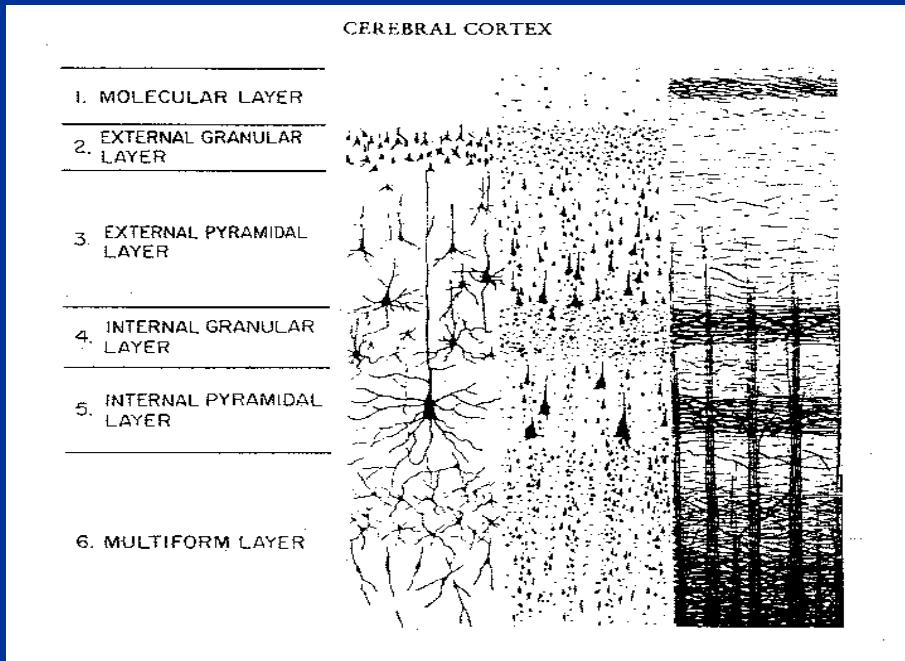
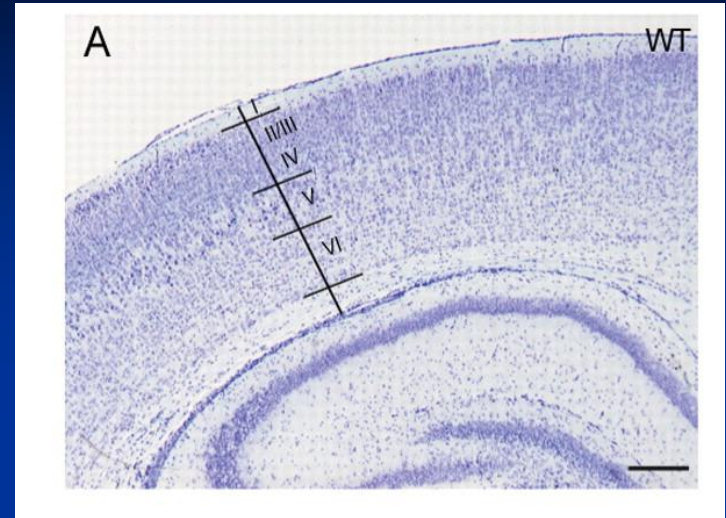
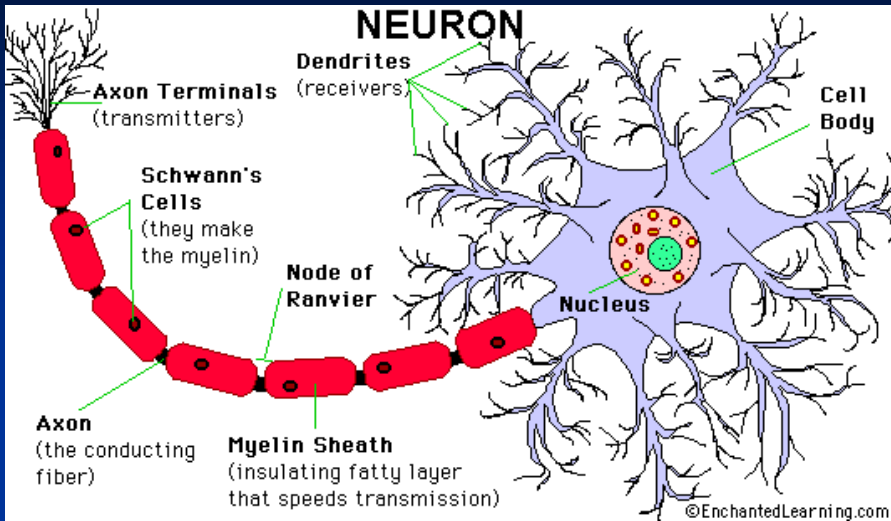
GABRB3(P11S) knockin: autism, CAE

GABRB3(D110N) knockin: IS

GABRB3(N120D) knockin: LGS

GABRG2(K328M) knockin :FS, SUDEP

# Neuron is very complicated and each neuron has many synapses to communicate with other neurons

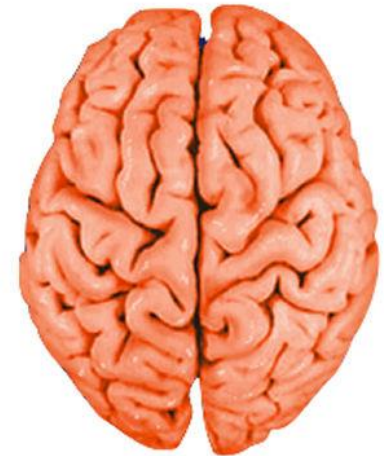


## Where is GAT-1 ??

Mouse brain

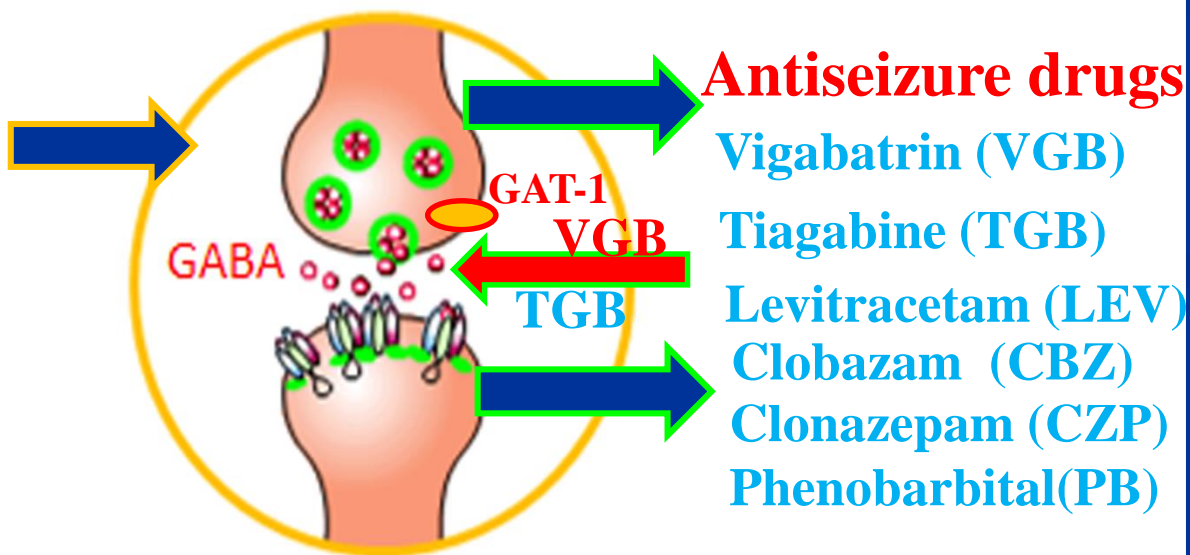
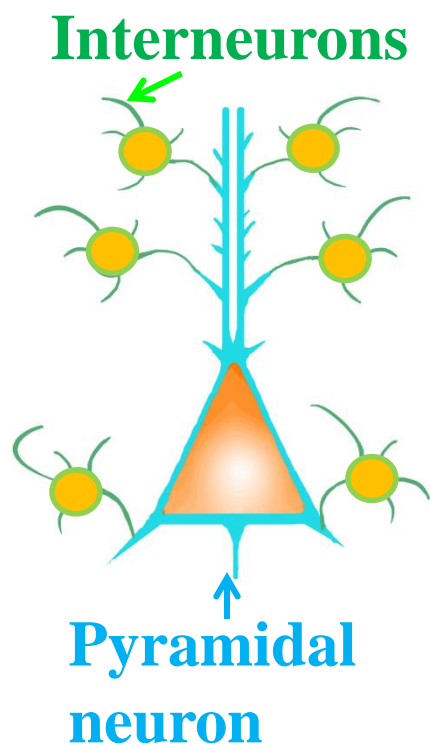


Human brain



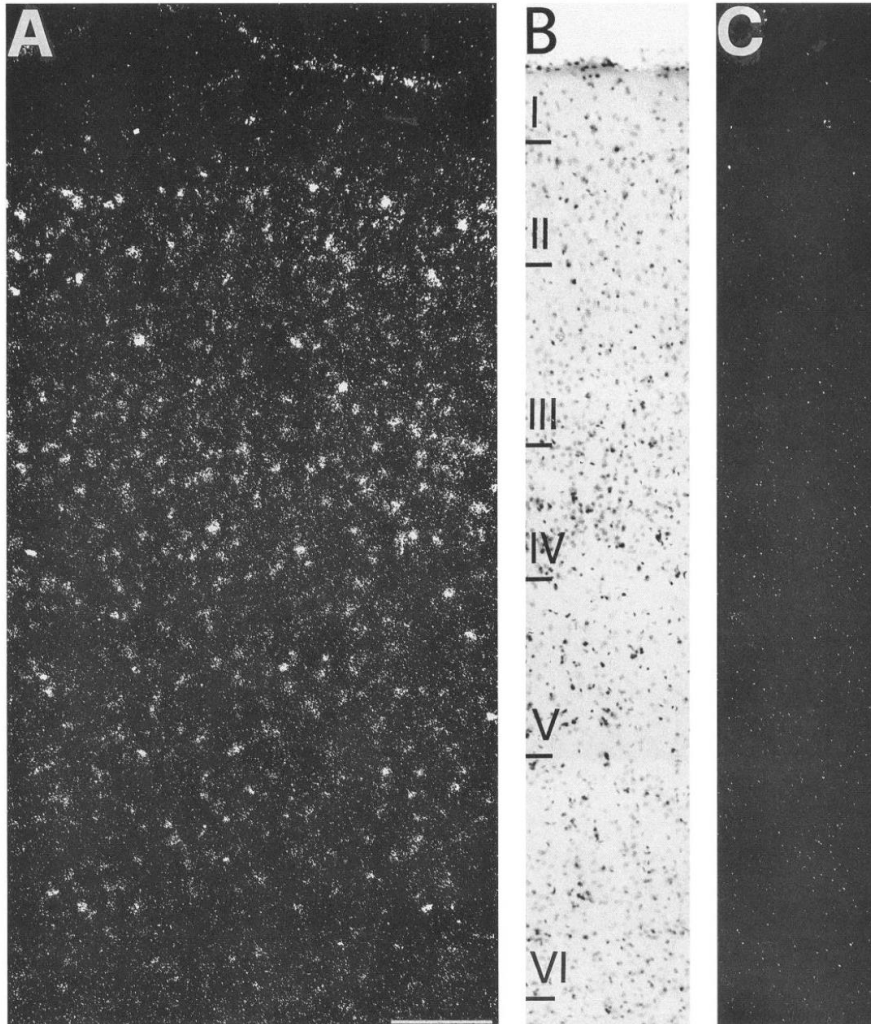
Our previous work has identified cortical GABAergic interneurons are autophagic in *Gabrg2*<sup>+/*Q390X*</sup> mice.

This is likely due to the accumulation of the  $\gamma 2$  subunits are accumulated in GABAergic interneurons.



# where is GAT-1 expressed?

7736 Minelli et al. • GAT-1 in the Cerebral Cortex



**GAT-1 transcripts in the first somatosensory cortex of the adult rat. The high level of hybridization signal in layer IV.**

**GAT-1 and GAD67 are often colocalized.**

# GAT-1 is also expressed in some pyramidal neurons

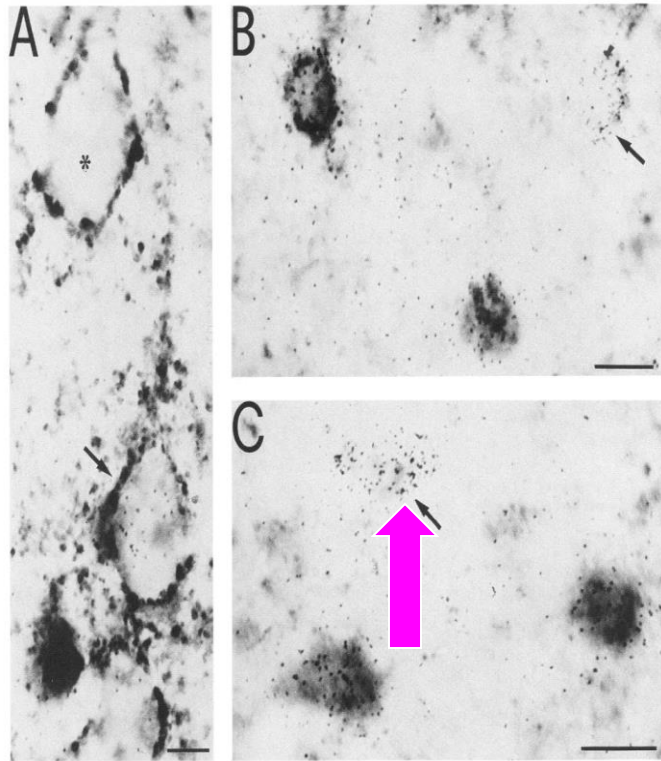


Figure 5. Simultaneous visualization of GAD67-ir and GAT-1 mRNA in rat cerebral cortex shows that all GAD67-positive cells express GAT-1 mRNA, but that not all GAT-1 mRNA expressing cells contain GAD67-ir (arrows). Of these, some are pyramidal neurons, as revealed by the dense clustering of GAD67-positive axon terminals on their somata and proximal dendrites (A). In A, asterisk indicates a pyramidal neuron not expressing GAT-1 mRNA. Exposure time: 15 d. A, Layer V; B and C, II. Scale bars: 20  $\mu$ m.

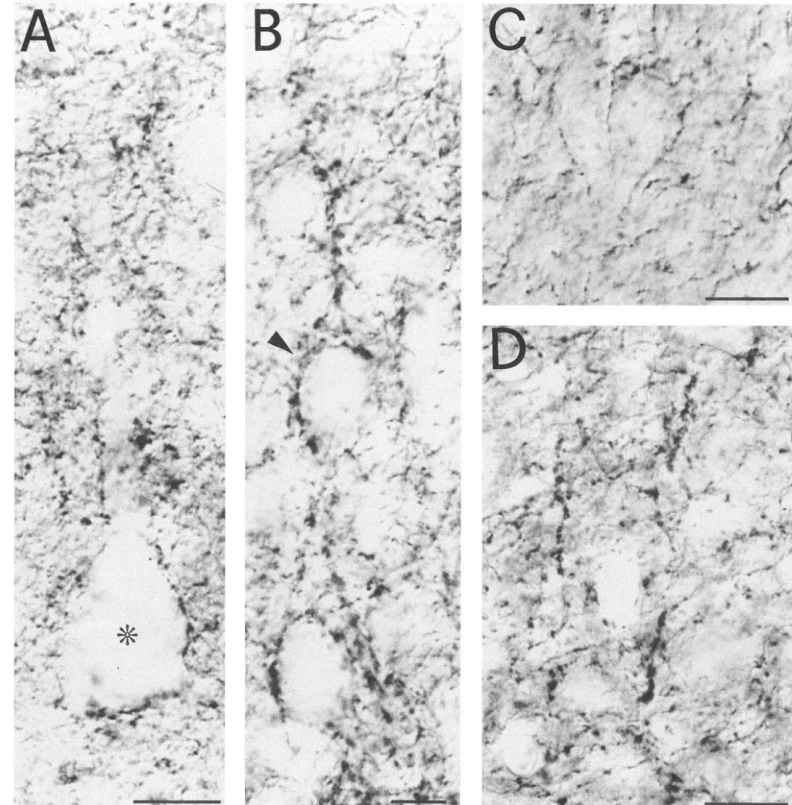
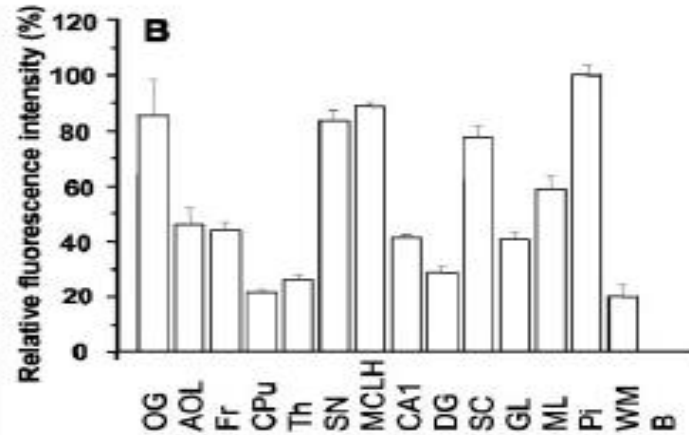
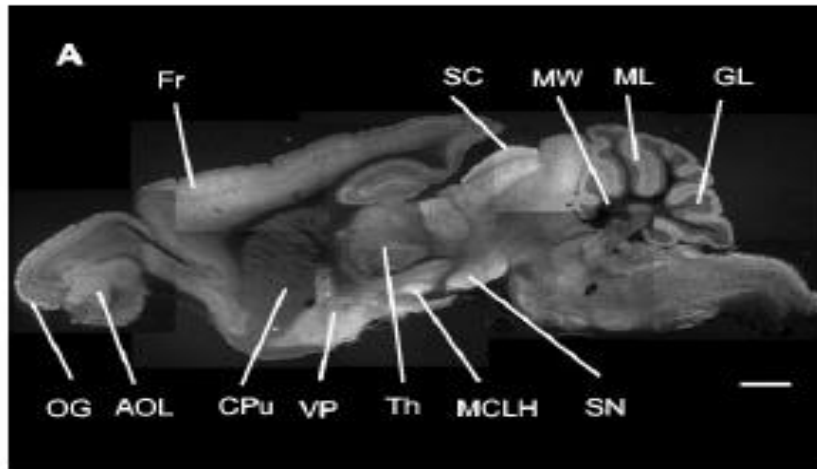


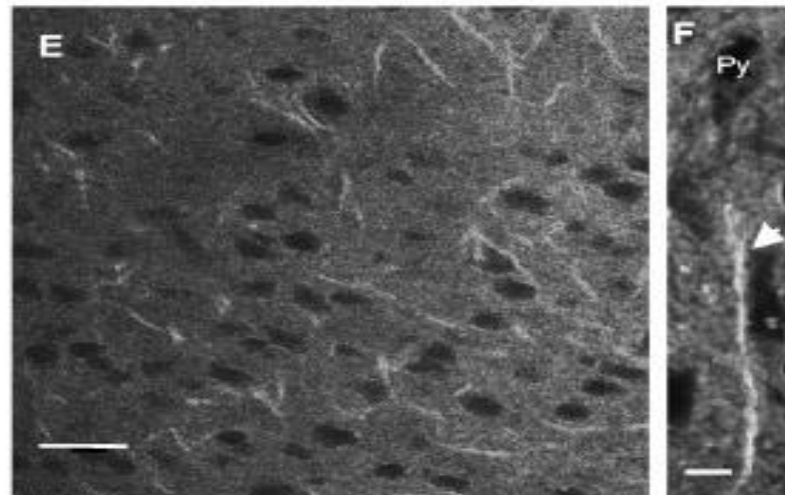
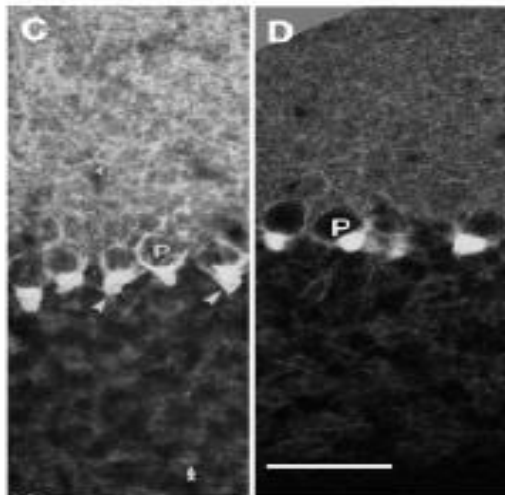
Figure 8. GAT-1 immunoreactivity within punctate structures that represent probable axon terminals (A and B) and in some fibers (C and D). The cell bodies and primary dendrites of both pyramidal (asterisk) and nonpyramidal (arrowhead) cells are outlined by stained axon terminals. Scale bars: A, 15  $\mu$ m; B, 10  $\mu$ m; C and D, 20  $\mu$ m.

# Number, Density, and Surface/Cytoplasmic Distribution of GABA Transporters at Presynaptic Structures of Knock-In Mice Carrying GABA Transporter Subtype 1–Green Fluorescent Protein Fusions

Chi-Sung Chiu,<sup>1</sup> Kimmo Jensen,<sup>4</sup> Irina Sokolova,<sup>1</sup> Dan Wang,<sup>3</sup> Ming Li,<sup>1</sup> Purnima Deshpande,<sup>1</sup> Norman Davidson,<sup>1</sup> Istvan Mody,<sup>4</sup> Michael W. Quick,<sup>3</sup> Stephen R. Quake,<sup>2</sup> and Henry A. Lester<sup>1</sup>



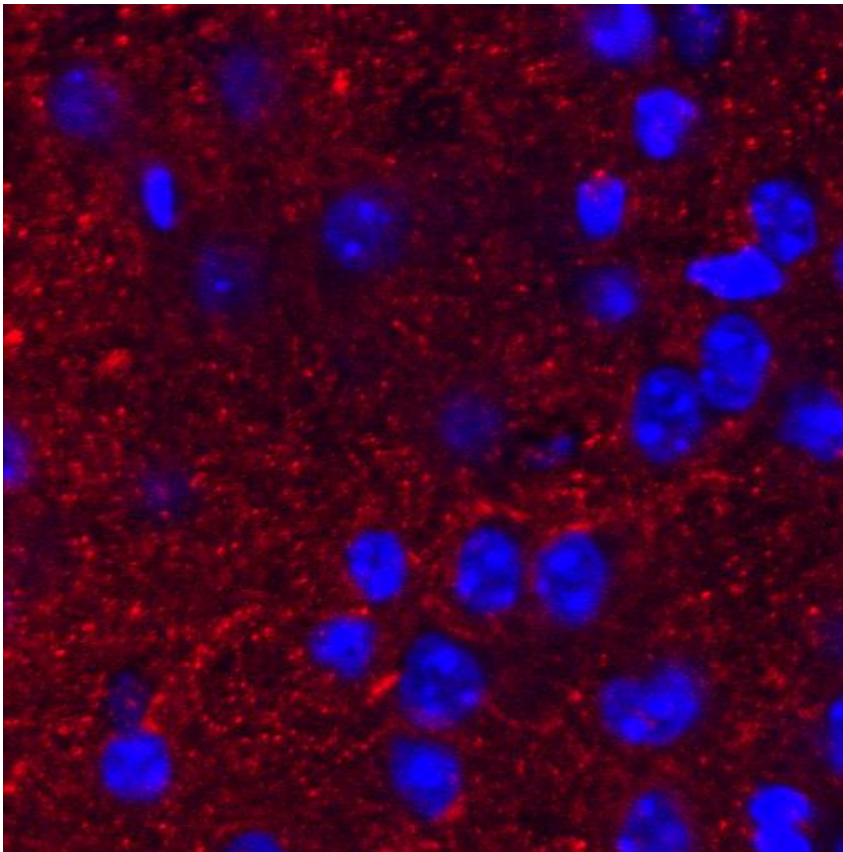
SN=substantia nigra  
 AOL=Anterior olfactory lobe  
 MCLH=Medial chiasmatic lateral hypothalamus  
 Pi=Pineaux



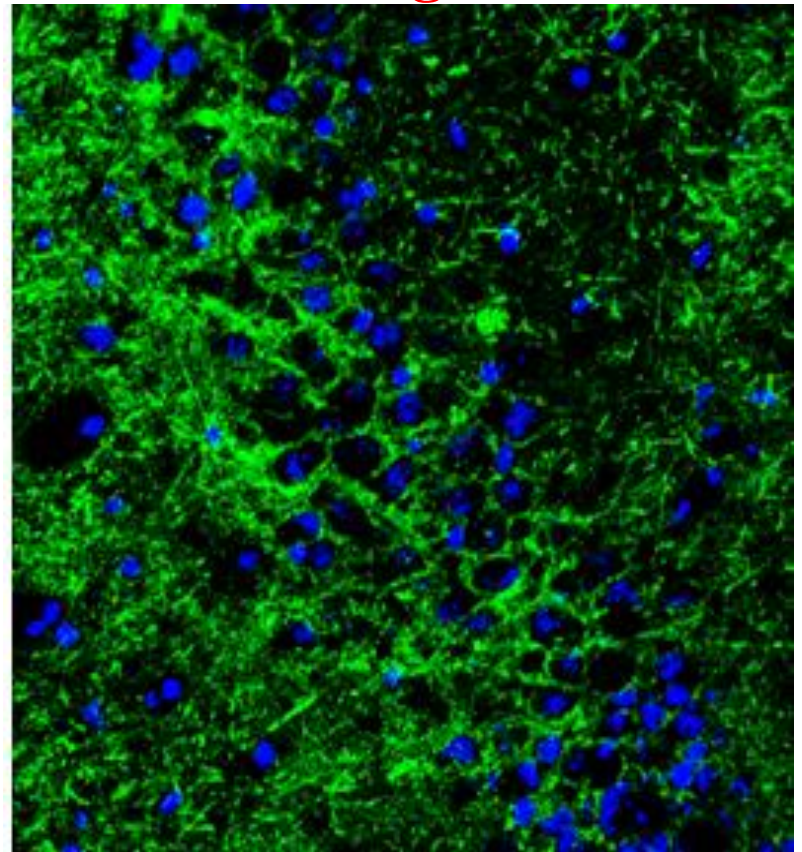
## Where is GABA transporter?

**GAT-1 immunoreactivity in cortex (red) and hippocampus (green)**

**3 months old mouse in C57/BL/6J mouse background**



**Cortex**



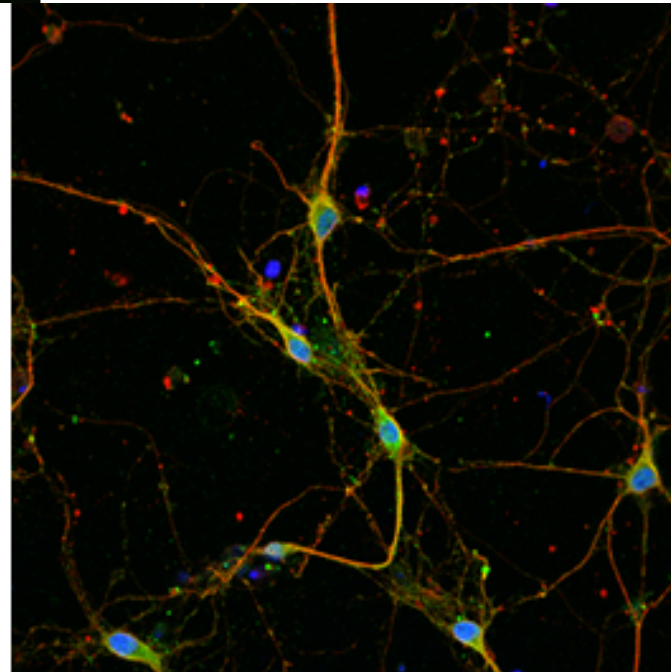
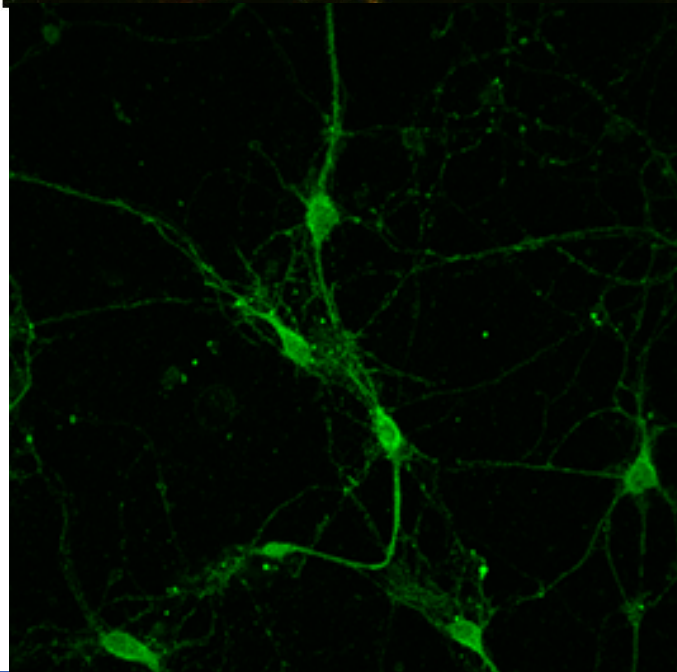
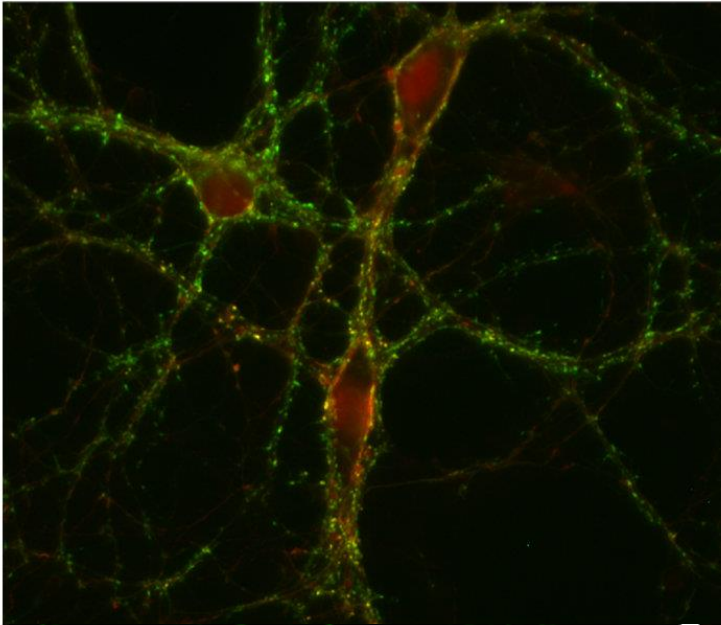
**Hippocampus**

**Mouse tissue**

At cell level

In cultured neurons

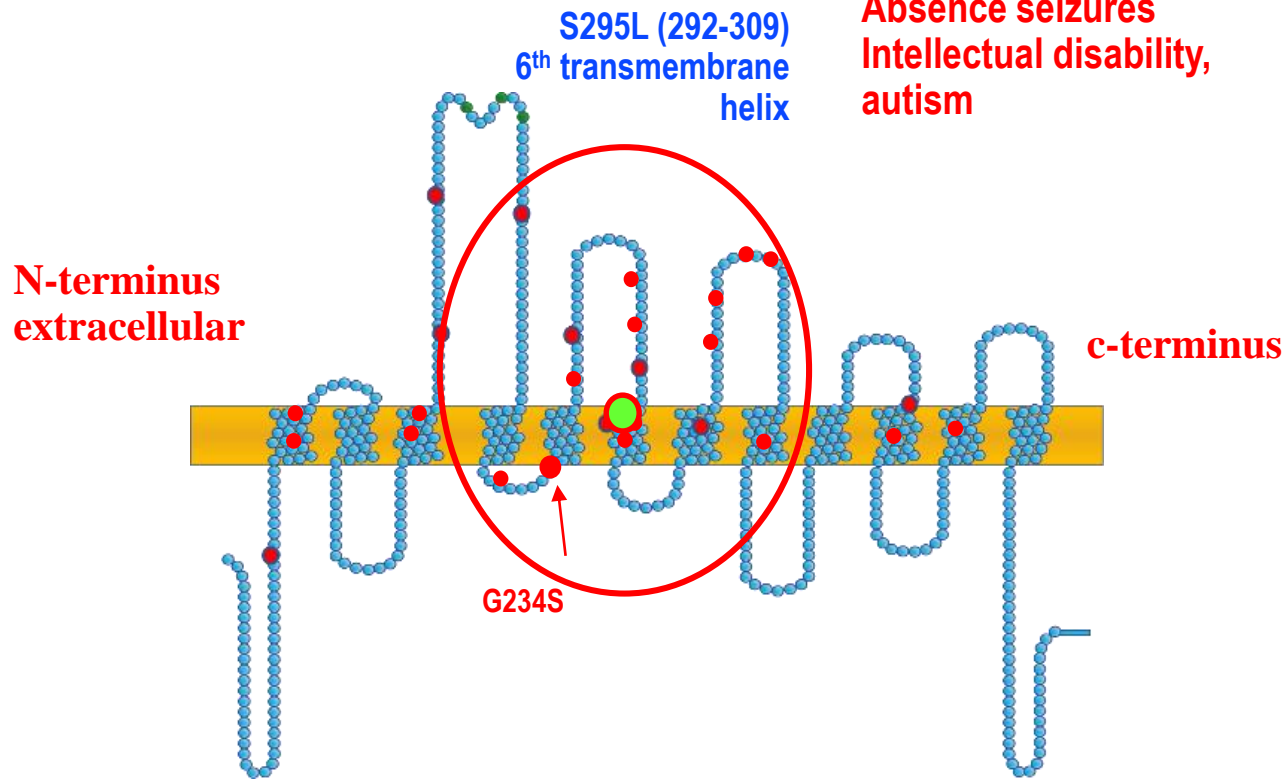
**GAT-1 immunoreactivity in  
Cultured neurons Green: GAT-1  
Red: Tubulin Tuji**



# Mutations in *SLC6A1* associated with epilepsy and intellectual disability

No. of *SLC6A1* (GAT-1) mutations=  
121

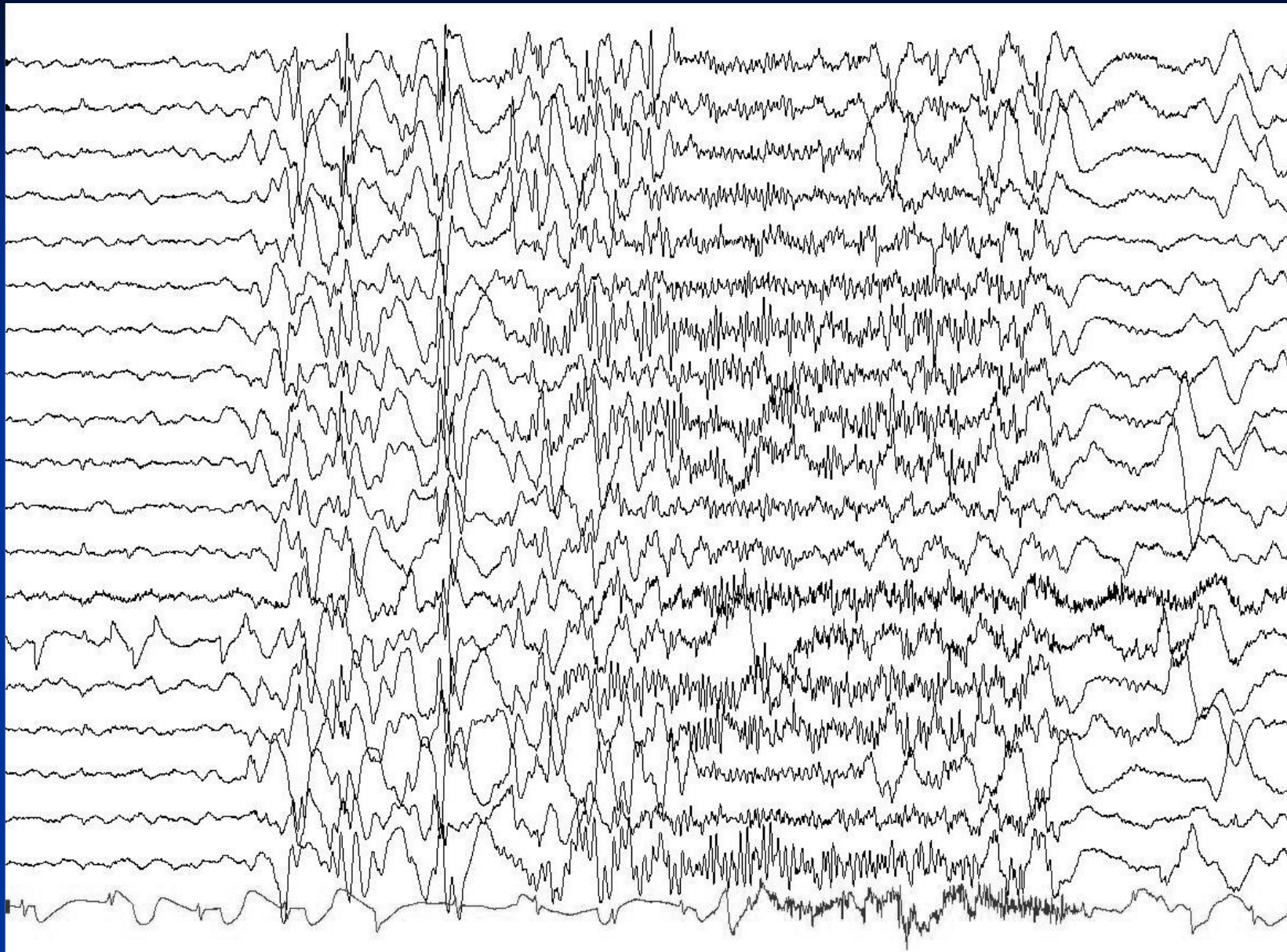
Phenotypes:  
Myoclonic atonic  
epilepsy  
Lennox-Gastaut  
syndrome  
Absence seizures  
Intellectual disability,  
autism



# EEG from the patient carrying SLC6A1(G234S) associated with LGS

A

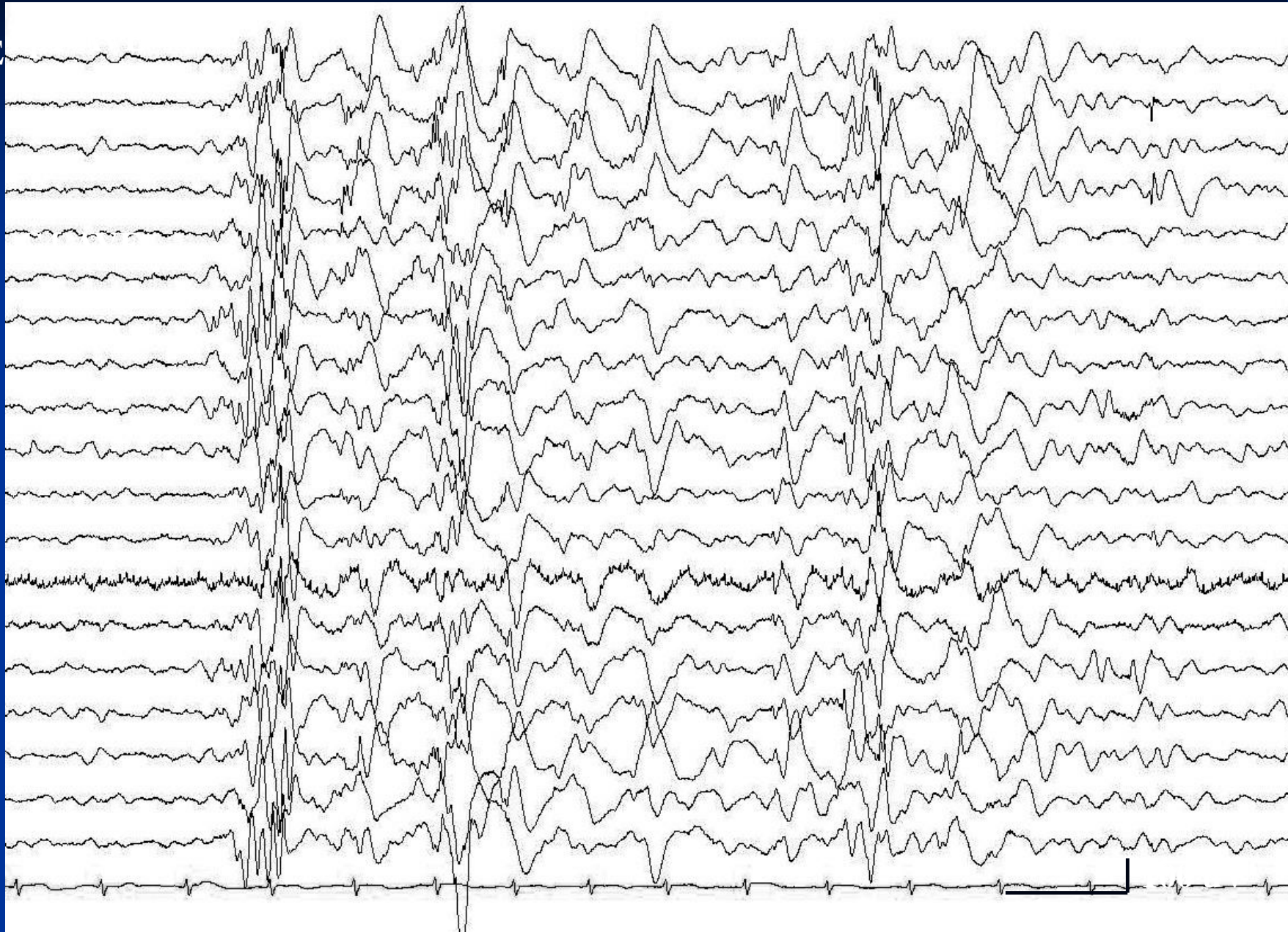
Fp1-AVE  
Fp2-AVE  
F3-AVE  
F4-AVE  
C3-AVE  
C4-AVE  
P3-AVE  
P4-AVE  
O1-AVE  
O2-AVE  
F7-AVE  
F8-AVE  
T3-AVE  
T4-AVE  
T5-AVE  
T6-AVE  
Fz-AVE  
Cz-AVE  
Pz-AVE  
EKG



100 uV  
1 sec

# EEG from the patient carrying SLC6A1(G234S) associated with LGS

fp2-AVE  
F3-AVE  
F4-AVE  
C3-AVE  
  
P3-AVE  
P4-AVE  
D1-AVE  
D2-AVE  
F7-AVE  
F8-AVE  
C3-AVE  
C4-AVE  
C5-AVE  
C6-AVE  
Cz-AVE  
Cz-AVE  
Pz-AVE  
EKG



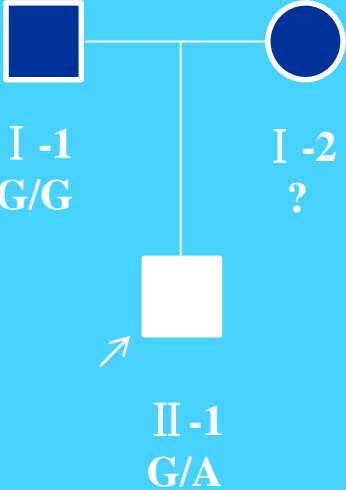
**Video showing the patient carrying SLC6A1(G234S) had tonic seizure**



# Molecular genetics of the patient

**A**

c.700G>A/p.G234S

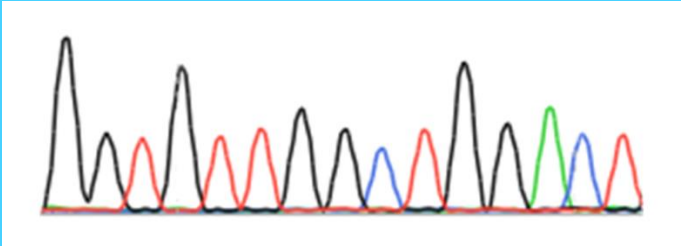


**B**

c.700G>A/p.G234S

G    V    **S**    W    T  
 ———  
 G G T G T T **A** G C T G G A C T

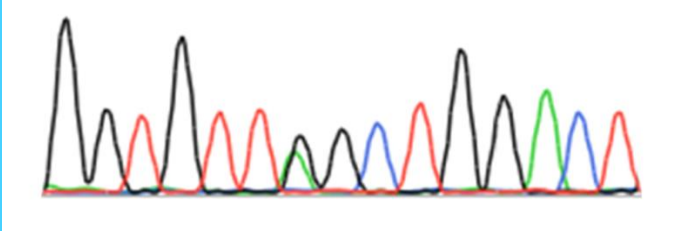
I -1



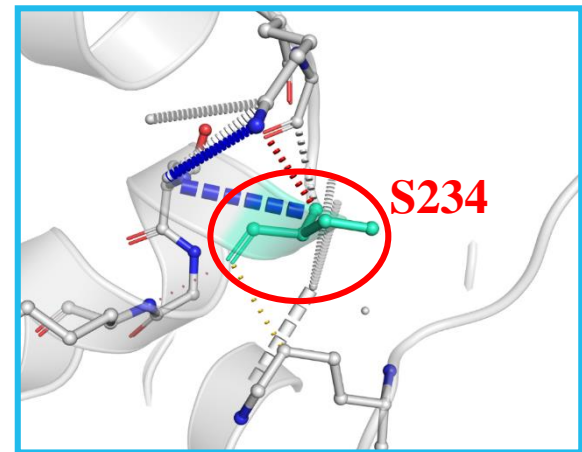
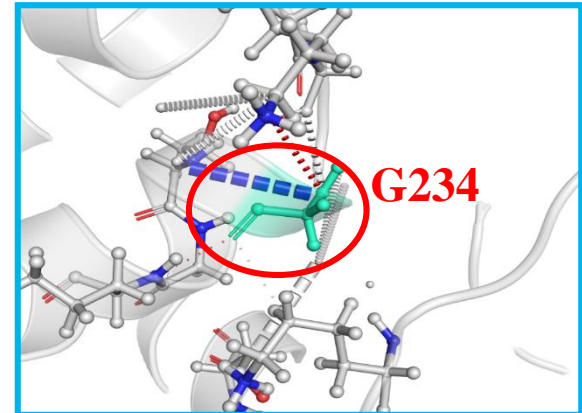
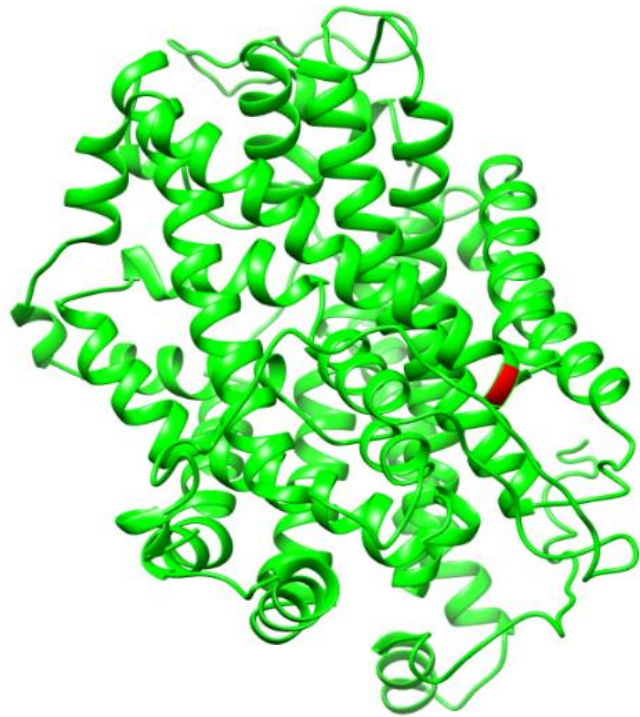
I -2

Not available

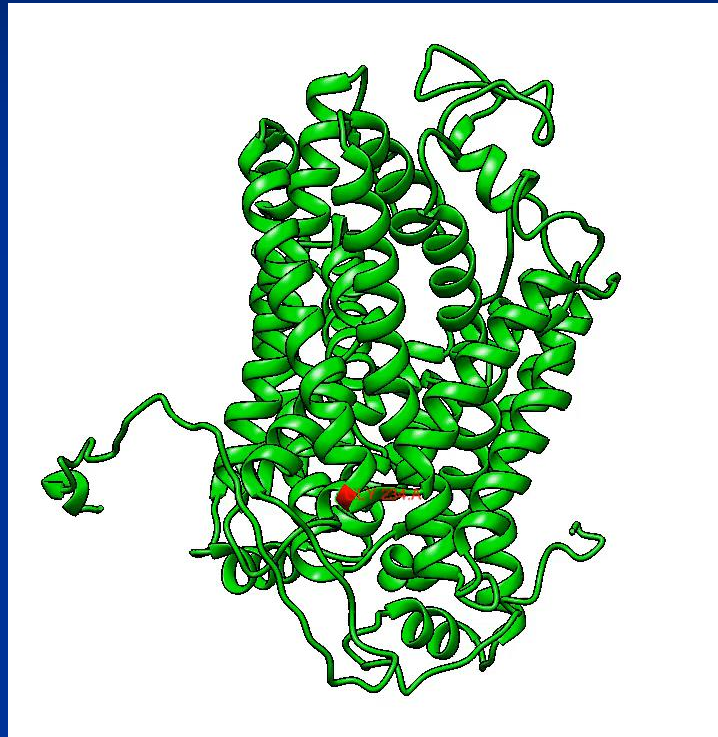
II -1



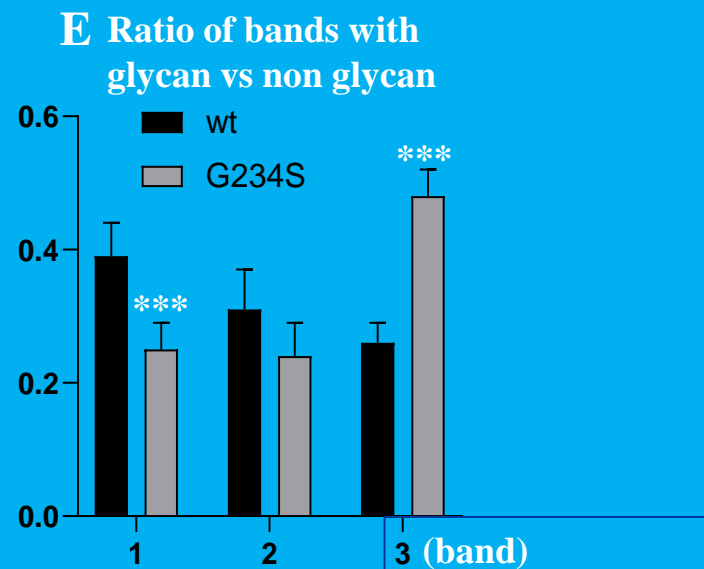
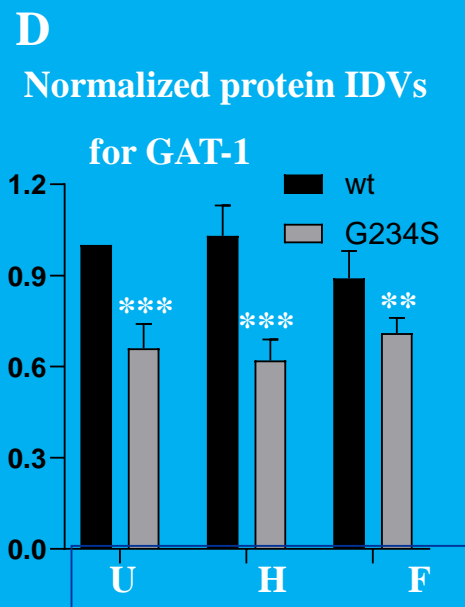
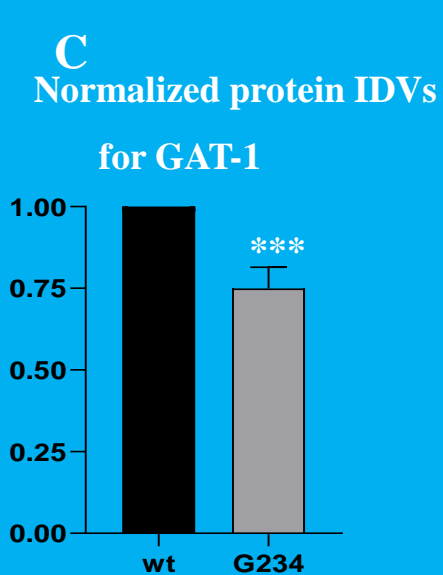
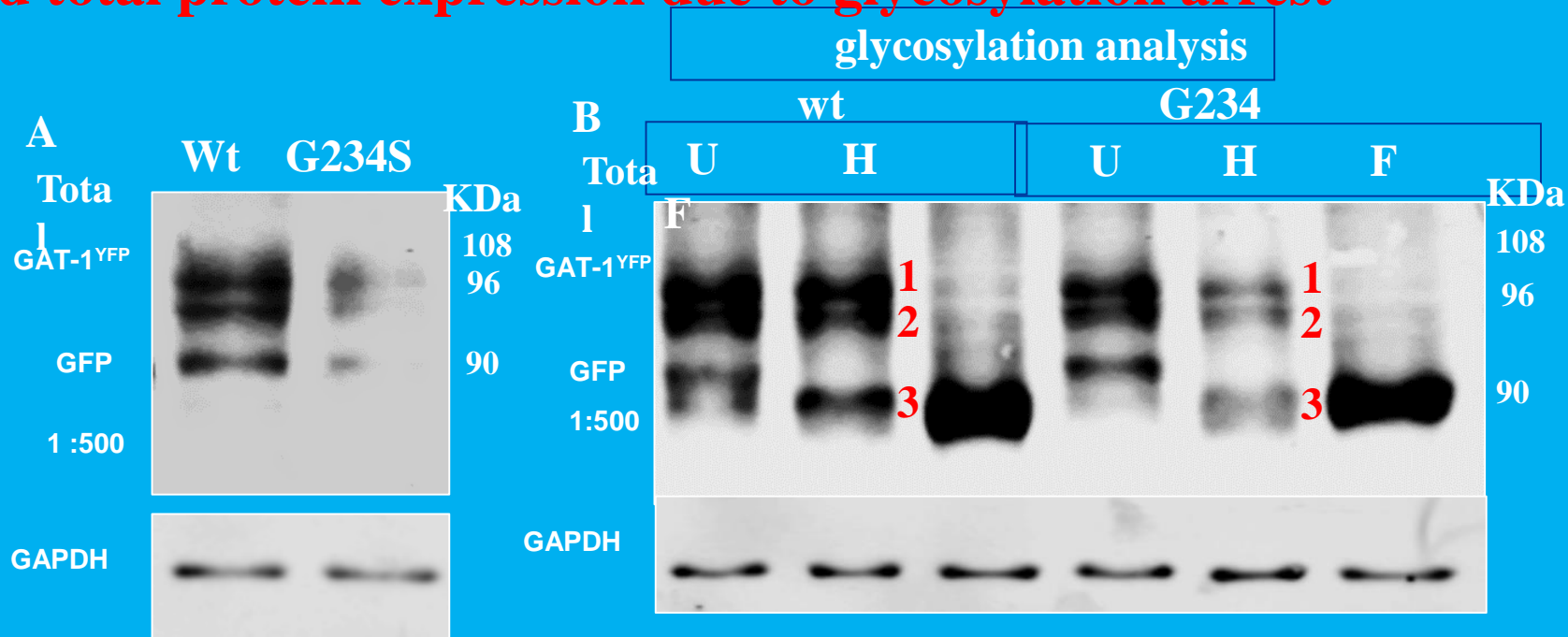
# Protein structural modeling and machine learning indicate the S234 destabilize the GAT-1 protein compared to G234



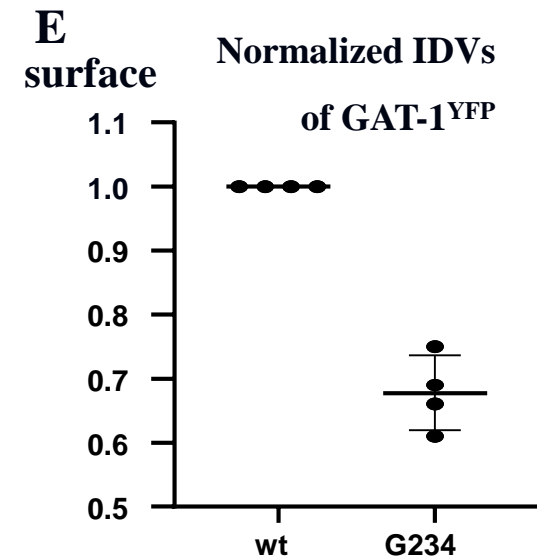
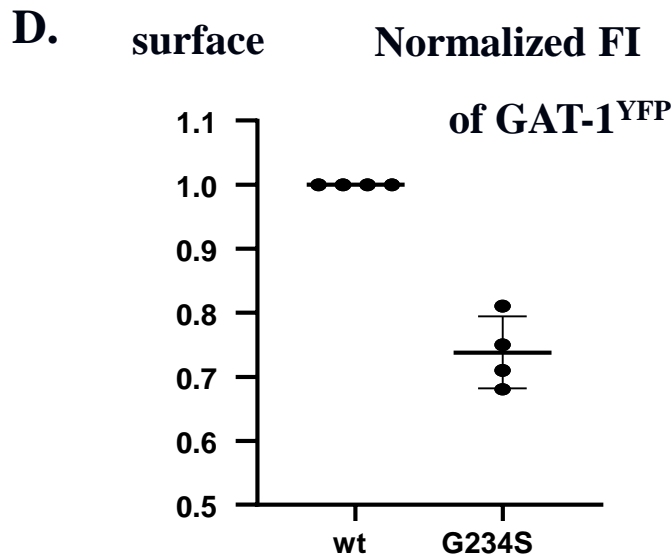
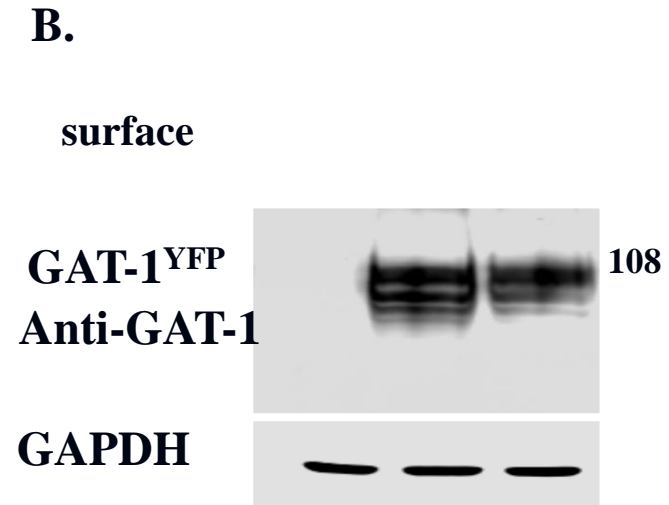
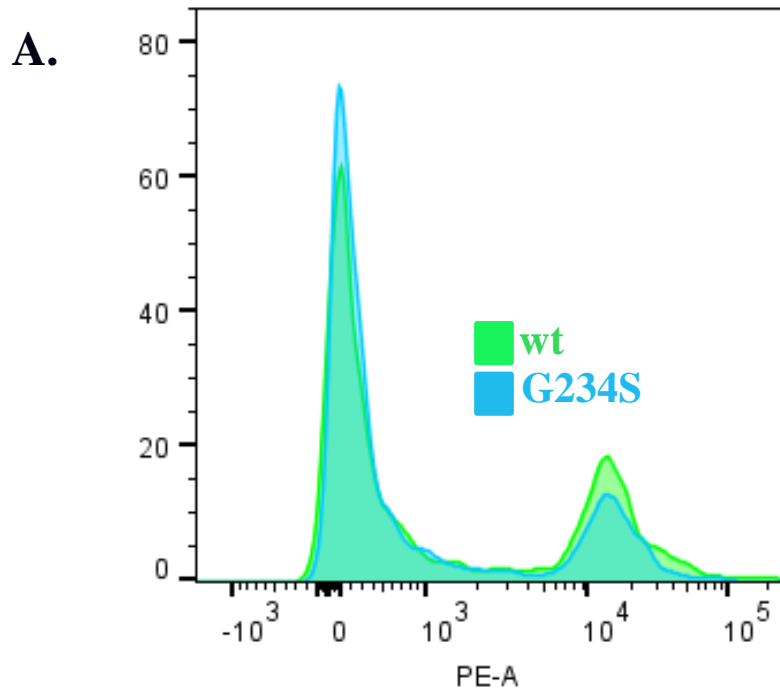
By Juexin Wang



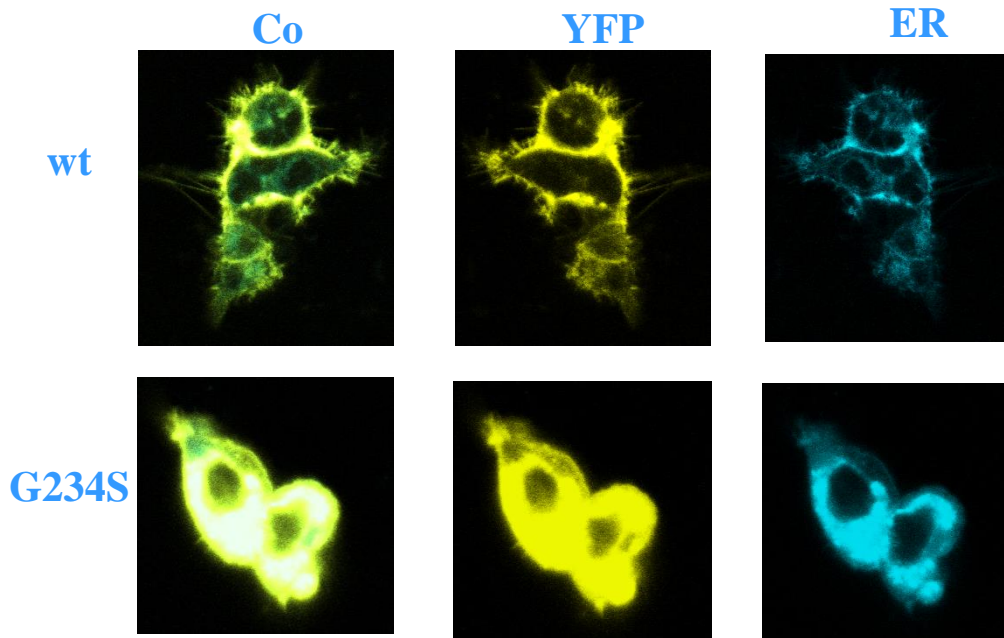
# Reduced total protein expression due to glycosylation arrest



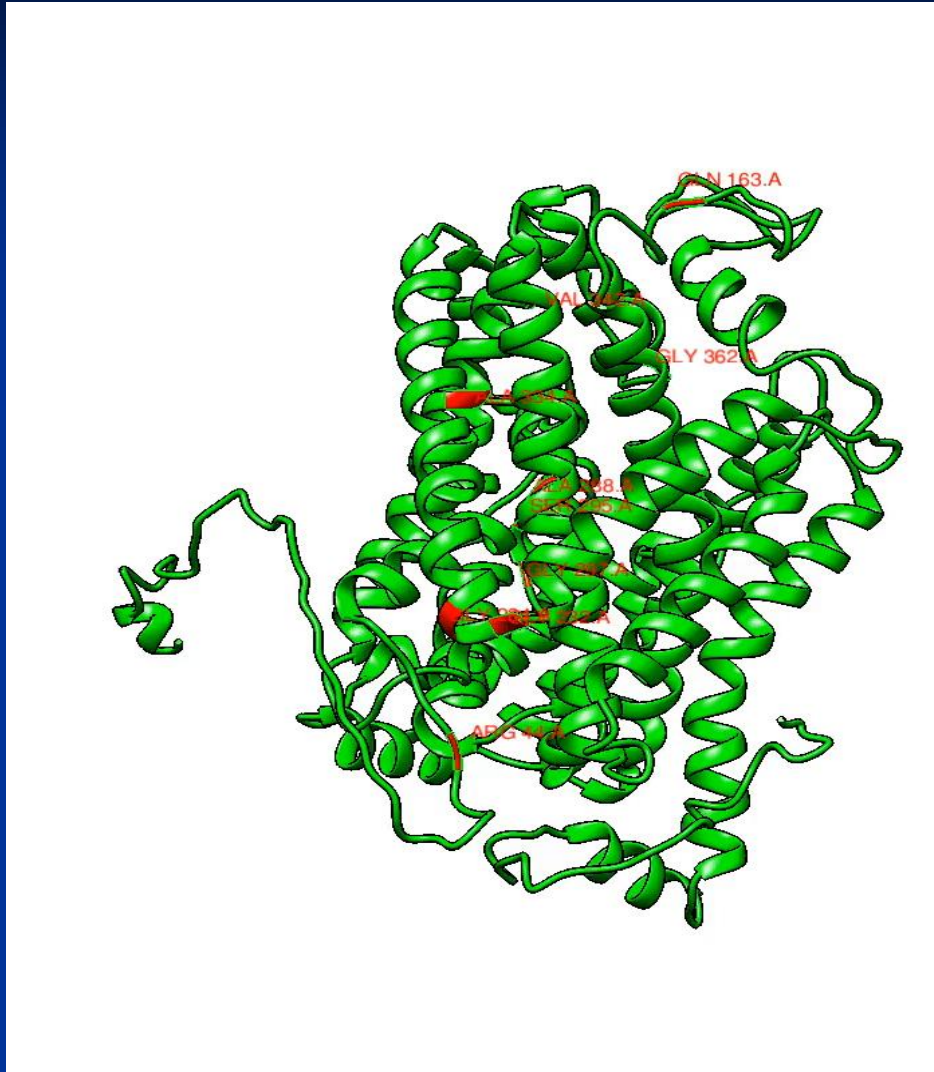
# Reduced surface protein expression as evaluated by flow cytometry and live cell surface biotinylation



# ER retention of the mutant GAT-1 (G234S) transporter



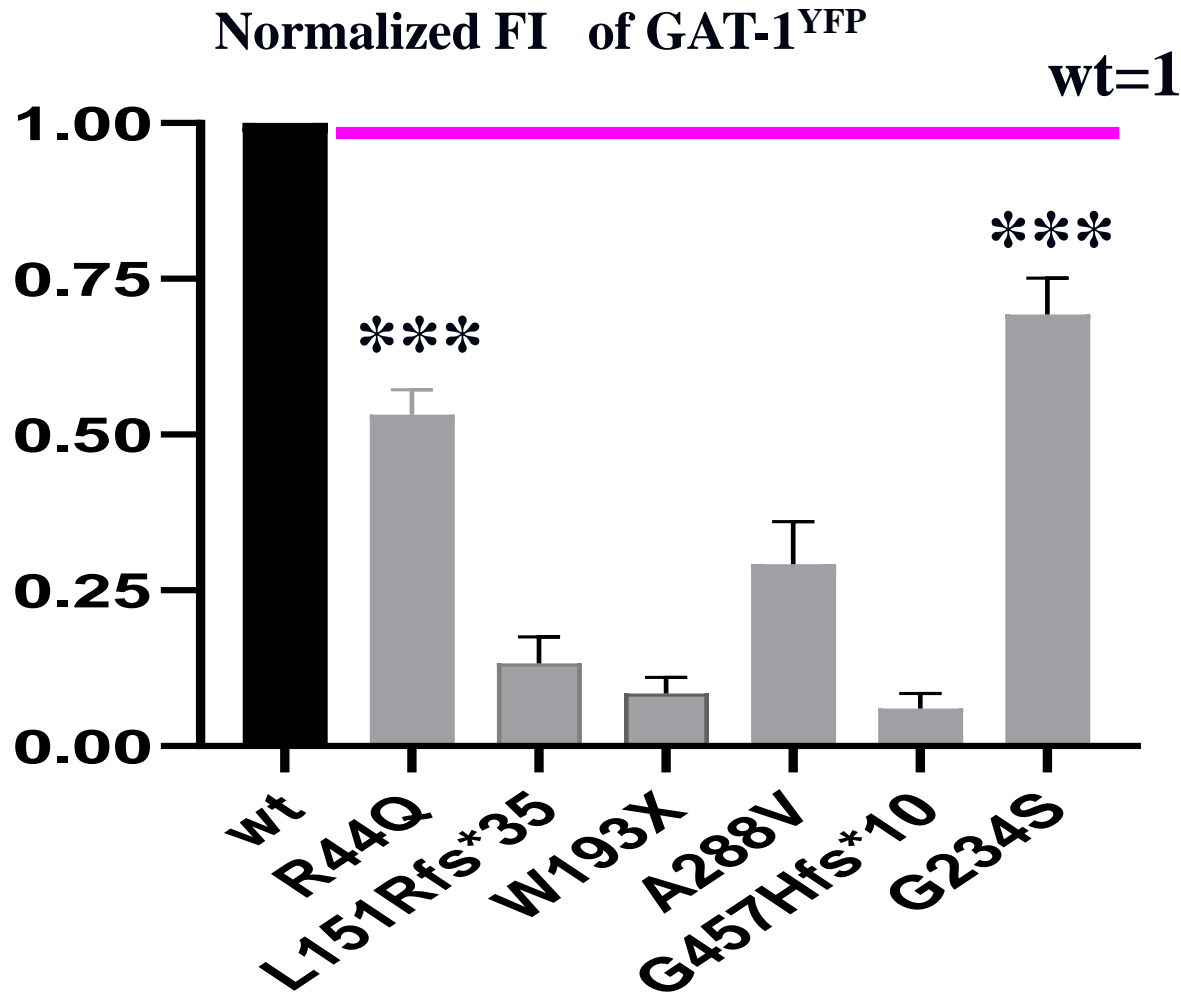
# Video showing multiple mutations in GAT-1



# Other mutations in *SLC6A1* associated with MAE and intellectual disability

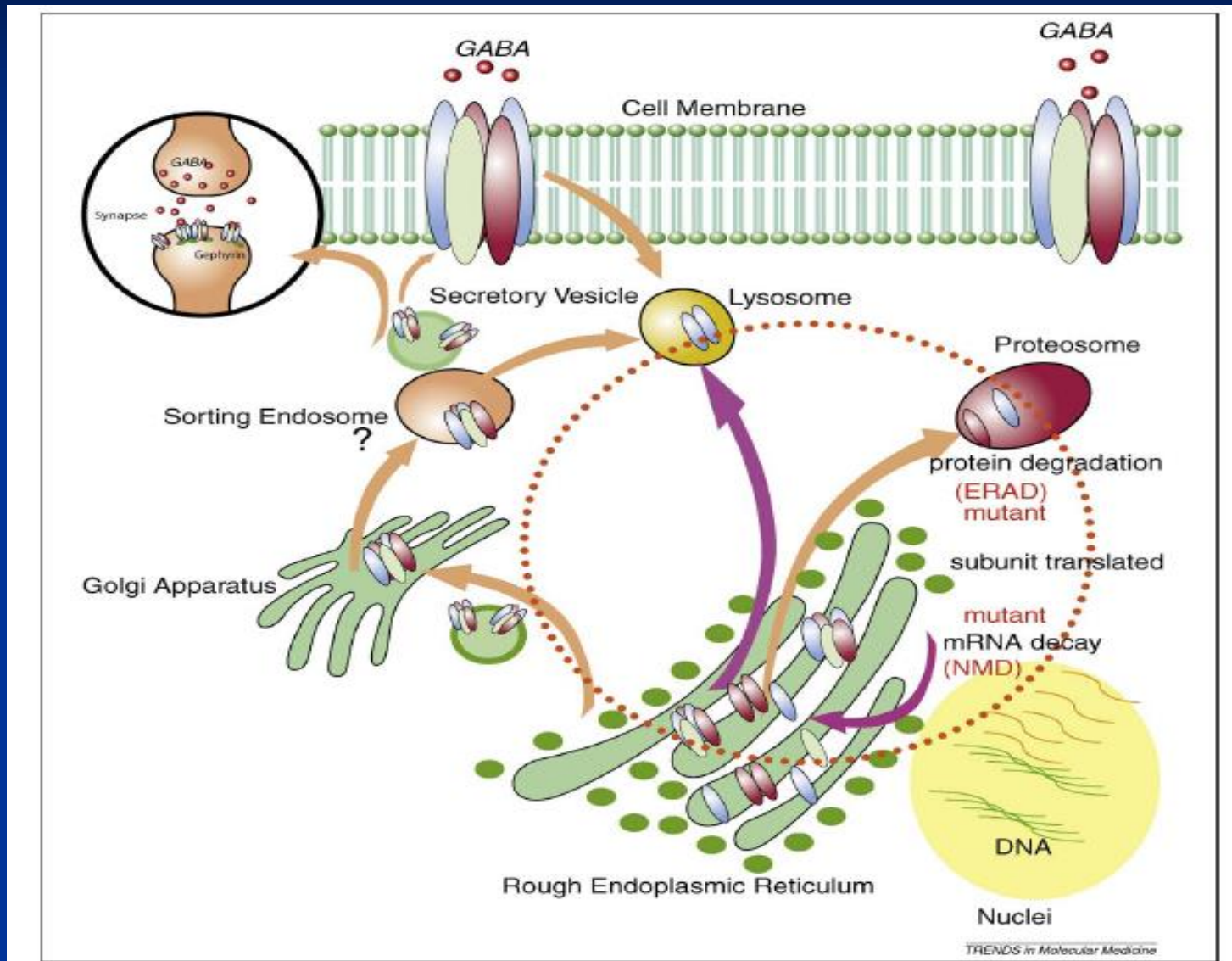


# Reduced surface GAT-1 protein expression is a common defect for SLC6A1 mutations



# The trafficking for mutant GAT-1 is unknown

1. Normal route of GABA receptor trafficking
2. NMD and ERAD pathways for the nonsense mutants





# Compare mouse models carrying mutations in *SLC6A1* and *GABR*

Mouse model:  
*SLC6A1*<sup>+/A288V</sup>

CRISPR/Cas9

mutant GAT-1



mutant GABR



*SLC6A1*<sup>+/A288V</sup>

*CAE, unclassified  
generalizedMAE, ID*

*GABRB3*<sup>+/N328D</sup>

*LGS*

# *Slc6a1*<sup>+/*A288V*</sup> mutant pups have better survival rate than *Gabrb3*<sup>+/*N328D*</sup> pups

**SLC6A1**<sup>+/*A288V*</sup>



**Gabrb3**<sup>+/*N328D*</sup>

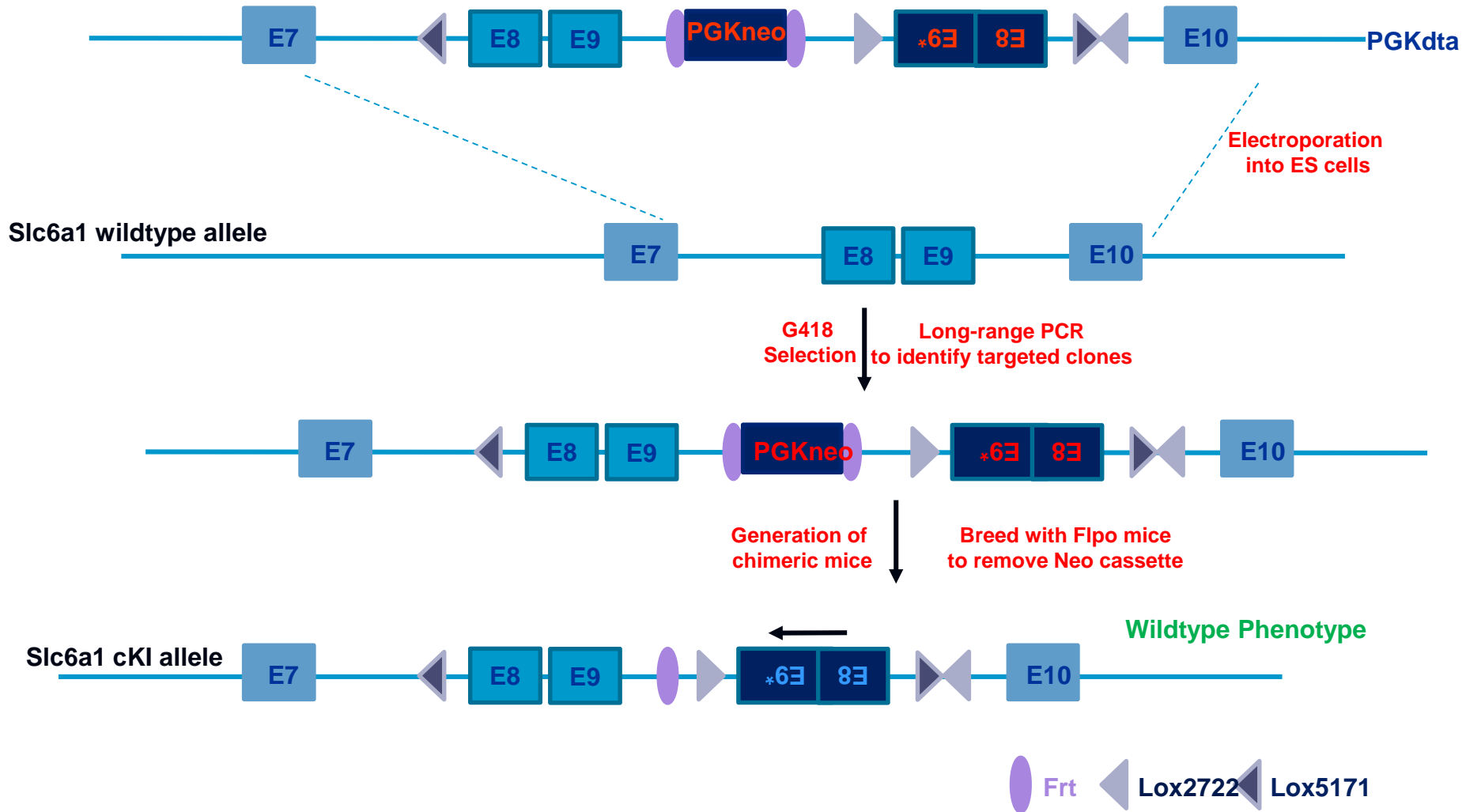


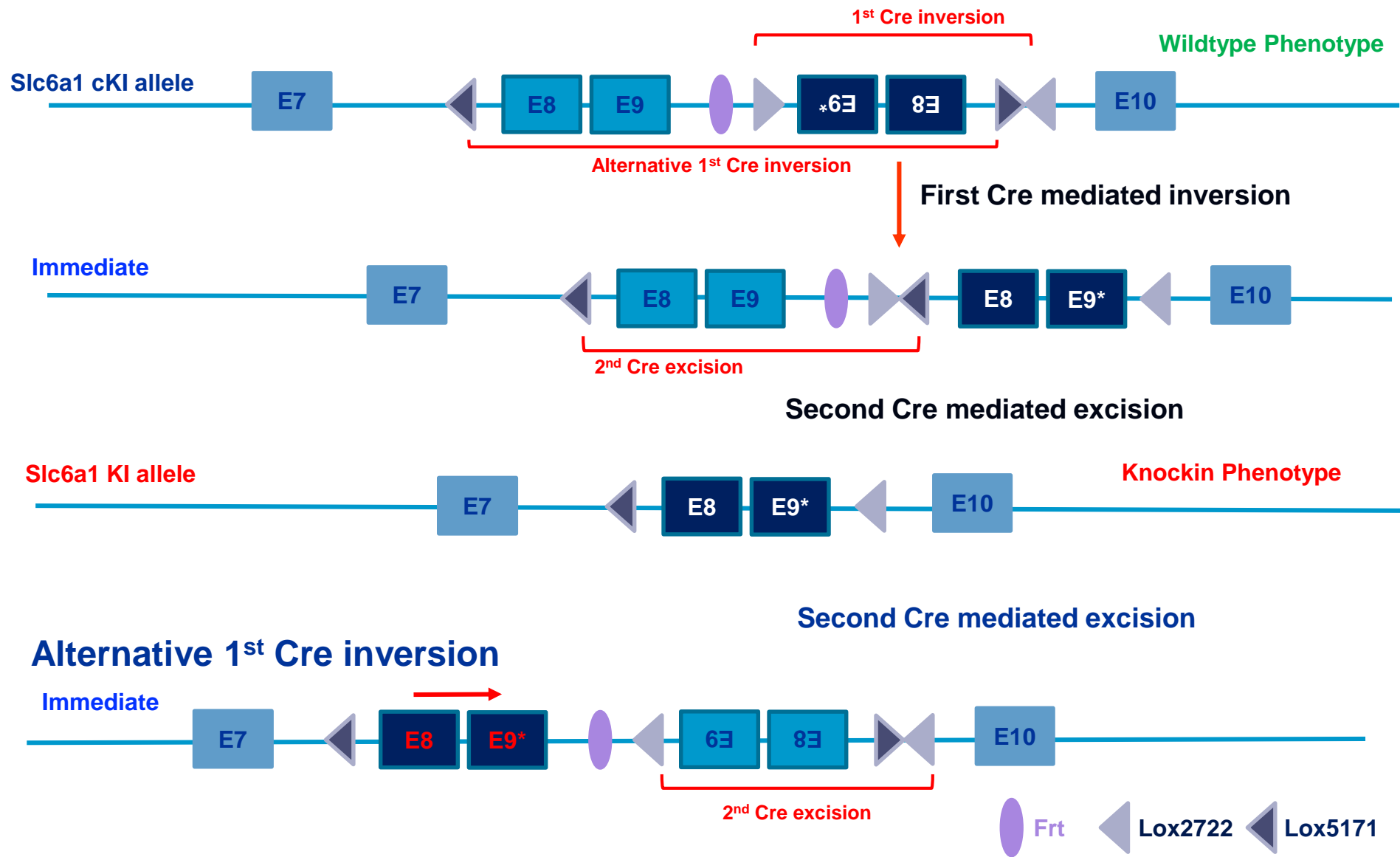
# mutant GABR



# Generation of Slc6a1 (X) Conditional Knockin Mice using FLeX Targeting Vector

X=A SLC6A1 mutation with different molecular defect than A288V mutation





## **Advantages and Features of Global Knockin:**

- **Efficient to generate using CRISPR mediated gene editing to introduce mutation directly into fertilized egg and therefore allow initial characterization of the effect of the mutation in a relatively short period of time.**
- **Mutation will be present in every cells of the animal and effects as soon as the gene is expressed during gestation.**

## **Advantages and Features of Conditional Knockin:**

- **More complicate, costly and longer time to generate.**
- **Mutation can be activated in a spatiotemporal (specific time and cell type) by breeding with tissue specific CreERT2 mice.**
- **More accurately reflected mutation occurs after conception or later in life in the human.**
- **Better tool to dissect effects of the mutation in a specific cell type or neurons, especially cell-to-cell interaction communication**

*With conditional knockin, we can specifically activate the mutation in neurons (different types of neurons) or astrocytes to characterize the disease phenotype.*

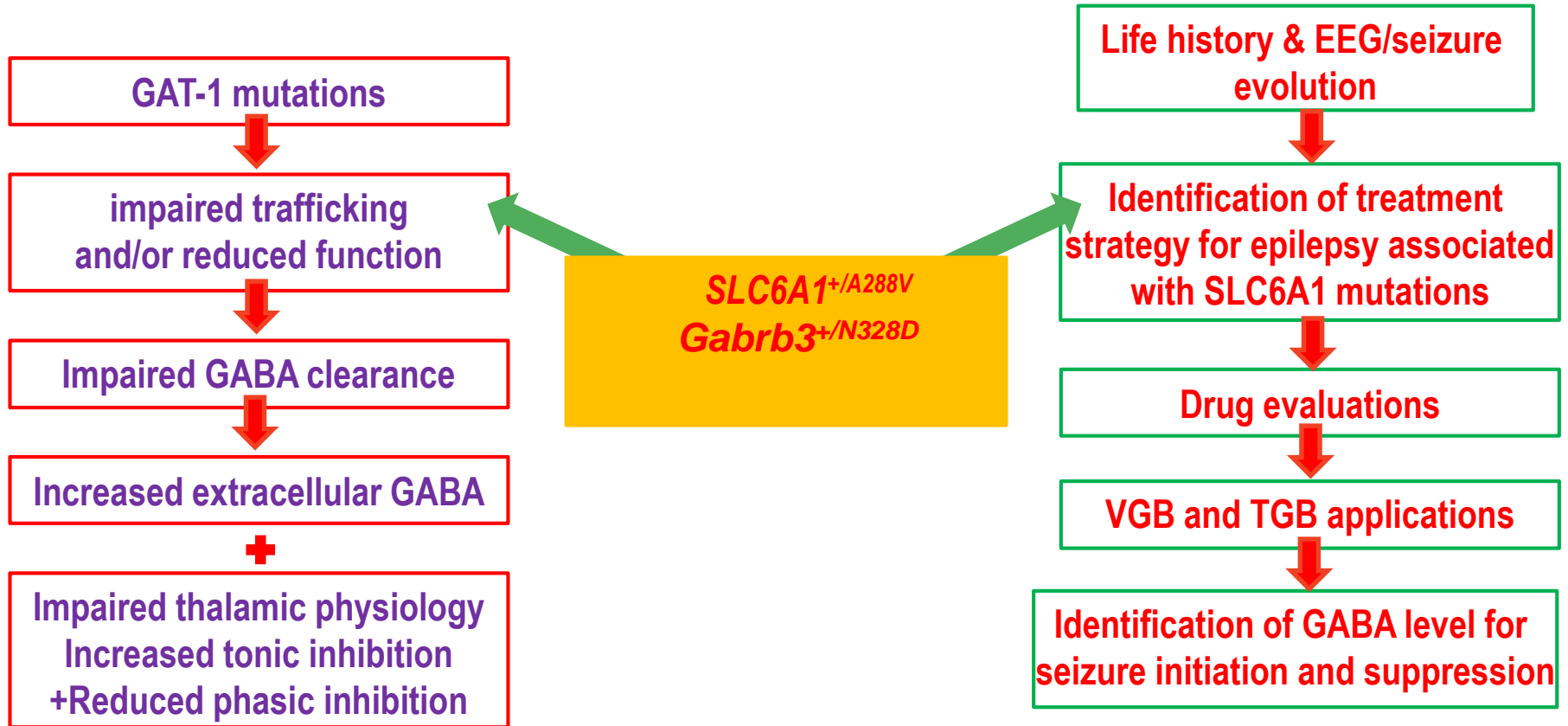
*We can compare the phenotypes in mouse with the phenotypes in human iPSC derived neurons or astrocytes.*

**Compared the effect of treatment with existing antiseizure drugs in mice carrying GAT-1(A288V) and GABRB3(N328D) mutations**

***In humans: Effective treatment by VPA and LTG in LGS caused by both GABRB3 and GAT-1***

***In SLC6A1<sup>+/A288V</sup> and Gabrb3<sup>+/N328D</sup> mice:????***

# Our Study Design

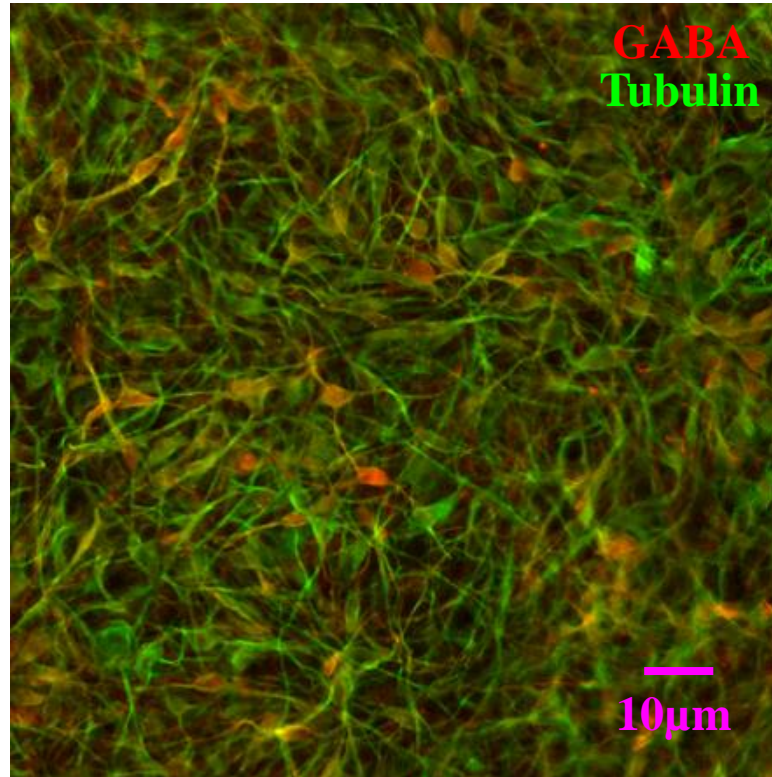


## Ongoing work:

**In addition to mouse models, we will validate the findings in human iPSC derived neurons and organoids**

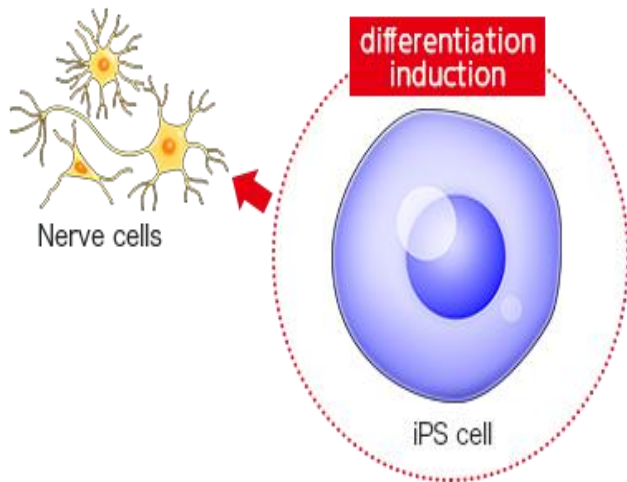
**GABAergic neurons derived  
from patient skin cells**

**GABAergic differentiated protocol (Maroof et al) was followed. At Day 26 post-plating cultures were fixed and immunostained for GABA (red) and Tuj beta-tubulin III (green).**



**Collaborator: Kevin Ess**

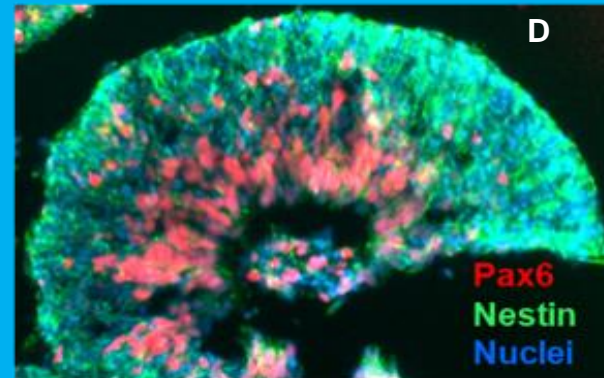
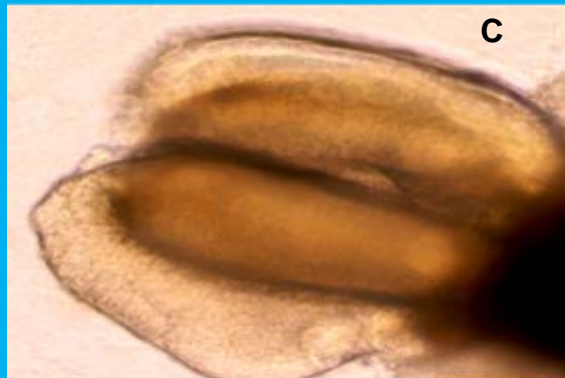
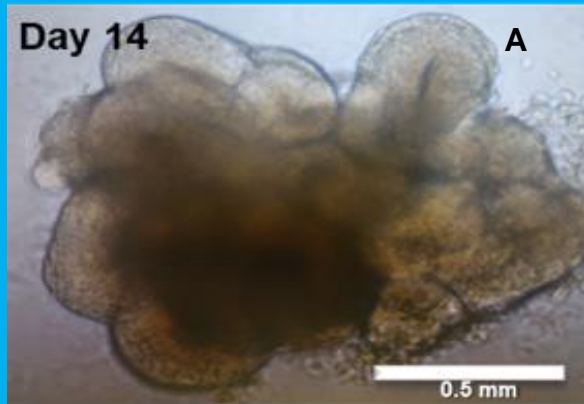
**Current ongoing work in lab: differentiation of GABAergic interneurons from An Angelman syndrome patient skin cells. The Angelman patient was caused by Deletion of UBE3a and GABRB3. We are comparing the data from cells and GABRB3 knockout mouse.**



**GABRB3 knockout**



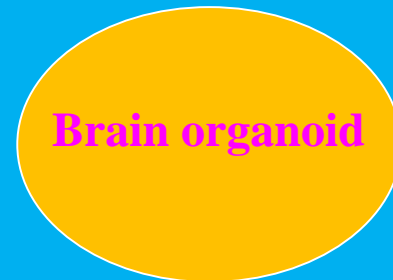
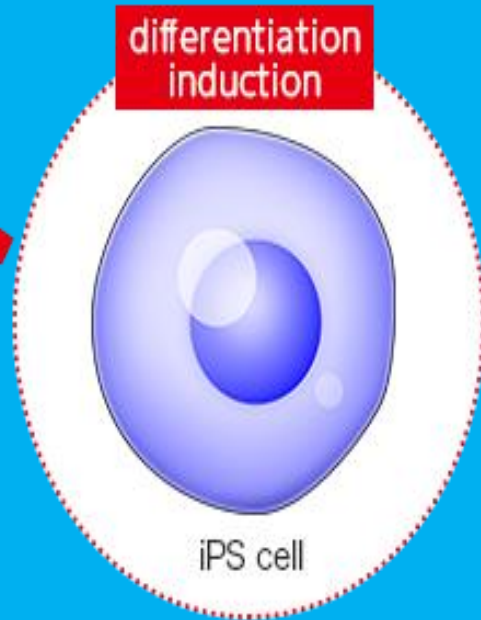
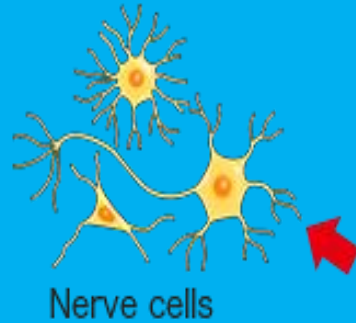
**Collaborator: Kevin Ess & Rob Carson**



**Generation of brain organoids from iPSCs in the Gama Laboratory (using modified protocols from Lancaster, 2016). At day 14 and 28, brightfield images show optically translucent neuroepithelial structures (A, B and C), while IHC staining (D) shows expression of the neural progenitor marker Pax6 (red) and the radial glia cell marker Nestin (green). Images are courtesy of Dr. Vivian Gama.**

**Collaborator:  
Vivian Gama**

Thanks to all the collaborators, we use multidisciplinary approaches to identify the underlying pathophysiology and mechanism-based therapies for mutations in *SLC6A1*



**Thanks for the invitation!**



**Musical city convention center in Nashville**

**End**